

Genetic drift does not sufficiently explain patterns of electric signal variation among populations of the mormyrid electric fish *Paramormyrops kingsleyae*

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Received June 24, 2015

Accepted February 20, 2020

Communication signals serve crucial survival and reproductive functions. In Gabon, the widely distributed mormyrid fish *Paramormyrops kingsleyae* emits an electric organ discharge (EOD) signal with a dual role in communication and electrolocation that exhibits remarkable variation: populations of *P. kingsleyae* have either biphasic or triphasic EODs, a feature that characterizes interspecific signal diversity among the *Paramormyrops* genus. We quantified variation in EODs of 327 *P. kingsleyae* from nine populations and compared it to genetic variation estimated from microsatellite loci. We found no correlation between electric signal and genetic distances, suggesting that EOD divergence cannot be explained by drift alone. An alternative hypothesis is that EOD differences are used for mate discrimination, which would require *P. kingsleyae* be capable of differentiating between divergent EOD waveforms. Using a habituation-dishabituation assay, we found that *P. kingsleyae* can discriminate between biphasic and triphasic EOD types. Nonetheless, patterns of genetic and electric organ morphology divergence provide evidence for hybridization between these signal types. Although reproductive isolation with respect to signal type is incomplete, our results suggest that EOD variation in *P. kingsleyae* could be a cue for assortative mating.

KEY WORDS: Animal communication, electric organ, electrocytes, genetic drift, signal evolution, weakly electric fish.

An overall goal in speciation research is to identify the evolutionary forces that generate divergence in populations, the targets of those evolutionary forces, and the relative order of the interactions that lead to speciation (Coyne and Orr, 2004). Rapidly speciating lineages are often characterized by highly diverse courtship signals (Diamond 1986; Allender et al. 2003; Mendelson and Shaw 2005; Boul et al. 2006; Mullen et al. 2007; Arnegard et al. 2010b). This has impressed on numerous evolutionary biologists the importance of behavioral isolation as a driving force in speciation, perhaps best summarized by the perspective of Mayr: “if we were to rank the various isolating mechanisms of animals according to their importance, we would have to place behavioral isolation far ahead of all others” (Mayr, 1963). Thus, it

is widely appreciated that differences in courtship signals are crucial in the *maintenance* of reproductive isolation between species. What is less clear is the relative importance of these differences in *initiating* speciation, and which evolutionary forces may act to produce these differences.

Communication signal differences can evolve due to selection or drift acting directly on signals or preferences for those signals, or alternatively, by selection to avoid maladaptive hybridization between populations divergent in other traits (Kirkpatrick and Ryan 1991). Contemporary patterns of phenotypic divergence within species radiations are most likely a combination of all these evolutionary mechanisms acting either separately, simultaneously, or sequentially (Glaubrecht 2010). It is a

fundamentally difficult problem to tease apart the contributions of the evolutionary forces that act on communication signals because of two important limitations. First is the ability in many systems to link *contemporary* forces acting in populations and the *historical* forces that have led to species diversity. Second, numerous communication system modalities are characterized by difficulty linking the anatomical and physiological substrates of signal generation and perception.

The first limitation can be circumvented in systems where *intraspecific* phenotypic variation parallels *interspecific* differences. This has been found in numerous adaptive radiations, such as in East African and Central American cichlids (Liem and Kaufmann 1984; West-Eberhard 1986; Meyer 1987; Maan et al. 2004), Darwin's finches (Smith 1987; Werner and Sherry 1987), *Heliconius* butterflies (Brower 1994), sticklebacks (Schluter and McPhail 1992), or coral reef hamlet fish (Picq et al. 2019). Intraspecific polymorphisms in traits that are characteristic of the interspecies diversity provide unique opportunities to decipher how evolutionary forces act in the early stages of divergence. The second limitation may be addressed by the study of model systems with comparatively simple physiological and anatomical substrates for signal generation and perception.

Electrocommunication in weakly electric fish offers an excellent opportunity to study the evolution of communication signals as electric signals are diverse, and easily measured and quantified. Most importantly, owing partly to the uniqueness of this sensory modality and the fact that weakly electric fish have served as a model system in neuroscience for more than 50 years (Lissmann 1958; Heiligenberg 1977), the anatomical and physiological basis of both perception and production of electric signals is well characterized and understood (Bennett and Grundfest 1961; Arnegard et al. 2010a; Gallant et al. 2011; Carlson and Gallant 2013). Weakly electric fish generate electric organ discharges (EODs) for orientation, navigation, and communication, and can sense perturbations in these self-generated signals through an array of electroreceptors distributed over their skin. In the species-rich African weakly electric fish (Mormyridae), a radiation within the *Paramormyrops* genus has given rise to more than 20 species over the last 0.5–2 million years in drainages of West-Central Africa (Sullivan et al. 2002; Sullivan et al. 2004; Lavoué et al. 2008). Species within this genus exhibit highly divergent EODs, which vary primarily in duration (0.5–8 ms) and in the number of phases present (Fig. 1A; Sullivan et al. 2000). P0-present EODs have three phases (triphasic) and are hypothesized to be the ancestral condition for *Paramormyrops* (Sullivan et al. 2004). They are characterized by the presence of a small, head negative prepotential (P0), which is absent in other species (termed P0-absent EODs, or biphasic EODs). Despite the subtle difference in EOD signals, the anatomical substrate for P0-present and P0-absent signals requires a considerable structural

reorganization of the cells that comprise the electric organ (electrocytes): individuals with triphasic EODs possess electrocytes with penetrating stalks and anterior innervation, whereas individuals with biphasic EODs, electrocytes have nonpenetrating stalks with posterior innervation.

Within the watersheds of Gabon, we recently described an intraspecific EOD polymorphism in the geographically widespread species *Paramormyrops kingsleyae*, whereby the majority of populations have triphasic EODs and a few have biphasic EODs (Fig. 2; Gallant et al. 2011). *Paramormyrops kingsleyae* thus reflects macroevolutionary patterns of EOD diversity across the *Paramormyrops* genus (Fig. 1B, Gallant et al. 2011), representing a “microcosm” of signal diversity and providing a great opportunity to study the evolutionary forces that initiate signal divergence. Among *Paramormyrops* species, playback experiments have revealed strong mating preferences for species-specific EODs (Hopkins and Bass 1981; Arnegard et al. 2006). In addition, Knollenorgan receptors, which mediate electrical communication in mormyrids, have been found to exhibit distinctive responses to different EOD waveforms (Arnegard et al. 2006). These findings indicate that individuals in the *Paramormyrops* species radiation are capable of detecting interspecific differences in EODs: is it also the case in the intraspecific polymorphism of *P. kingsleyae*?

Here, we assess evidence for neutral versus selective divergence of EOD properties in populations of *Paramormyrops kingsleyae*, in an ongoing effort to determine the evolutionary mechanisms that shape early divergence in EODs, the key variable trait within the *Paramormyrops* radiation and between mormyrids. This intraspecific polymorphism allows us to connect population-level processes to macroevolutionary patterns of diversity.

Gallant et al. (2011) considered drift as the most likely force to explain the observed variation in *P. kingsleyae* EODs based on three major findings: (1) evidence of substantial genetic partitioning between allopatric populations of *P. kingsleyae* (Arnegard et al. (2005); (2) clinal variation patterns in EOD duration and P0 magnitude across Gabon; (3) the discovery of morphologically “intermediate” individuals in Apassa and Bambomo Creek, where both biphasic and triphasic signal types co-occur, which were putatively considered as evidence of hybridization, suggesting that mating may not be fully assortative with respect to signal type. Taken together, these results suggest that signal divergence may increase with geographic distance, that mating may be random with respect to signal type in sympatric populations, and that allopatric *P. kingsleyae* populations are genetically isolated, patterns that are expected when drift is important in driving signal divergence (Wilkins et al. 2013). Whether genetic differentiation is accompanied by proportional levels of signal variation in this species is still unknown.

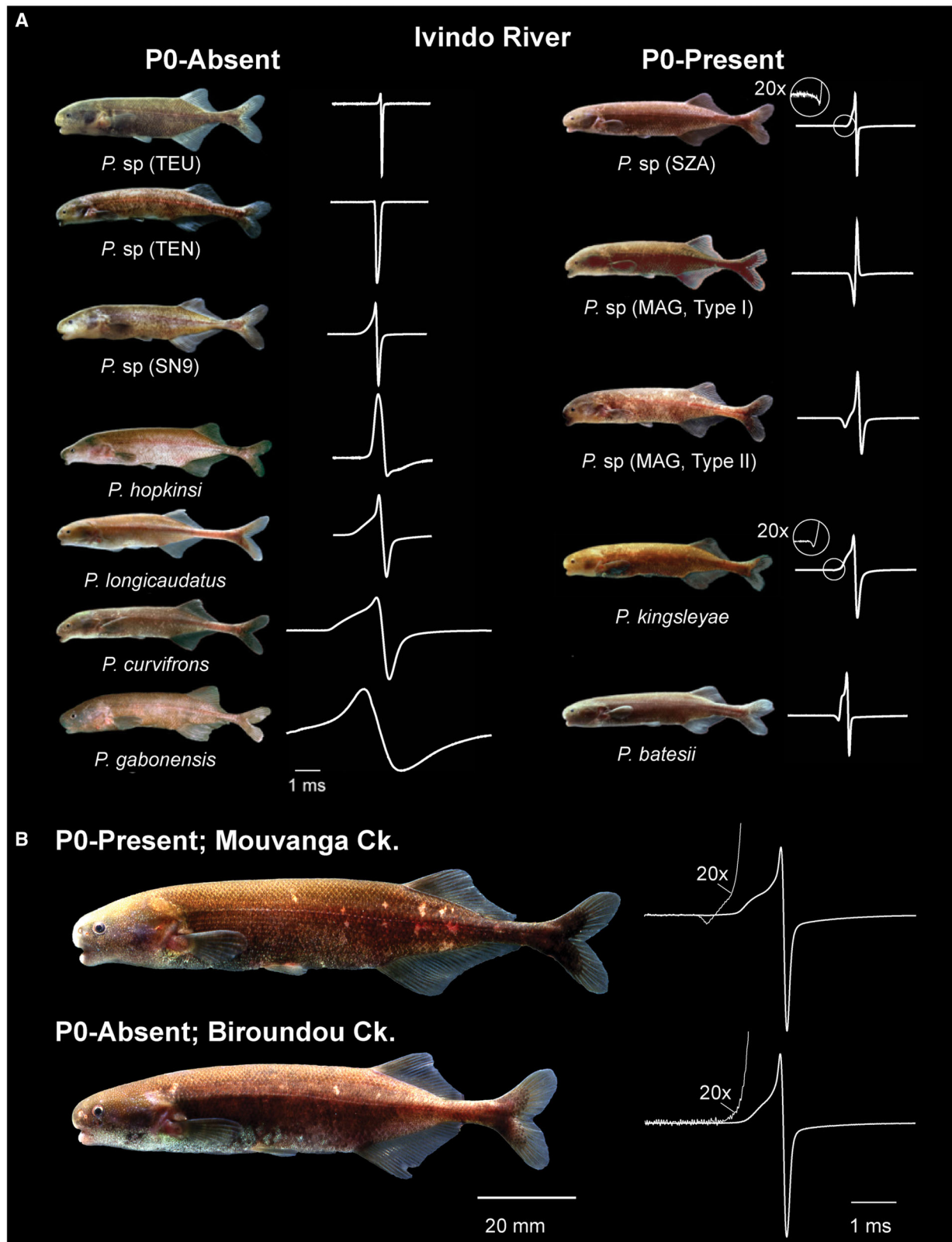


Figure 1. *Paramormyrops kingsleyae* EOD variation is a microcosm of EOD variation in *Paramormyrops*. A. 12 sympatric *Paramormyrops* specimens captured from the same locality in the Ivindo River illustrate EOD variation within the *Paramormyrops* genus in terms of duration and polarity, as well as in the number of phases. All *Paramormyrops* on the left have biphasic (P0-absent EOD) waveforms and all *Paramormyrops* on the right have triphasic (P0-present) EOD waveforms. B. *P. kingsleyae* EODs are variable in the presence of a small head negative phase (P0), see Gallant et al. (2011).

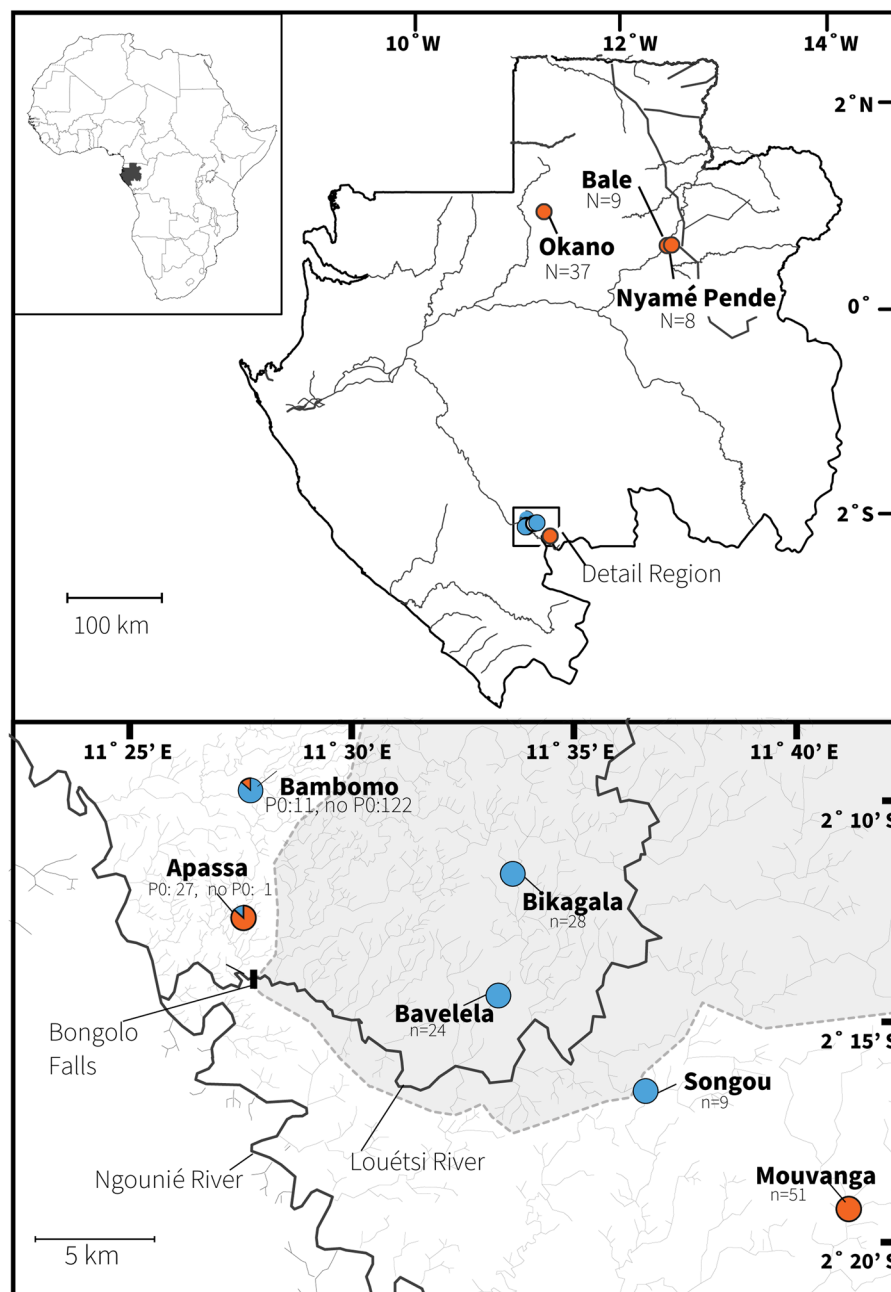


Figure 2. Map of study populations. Top map shows relationship between populations across Gabon. Box in the south indicates the location of the bottom, higher detail map, which shows the relationship of Southern populations near the confluence of the Louétsi and Ngounié rivers. For all maps, populations are indicated by pie charts, as reported in Gallant et al. (2011). Orange indicates proportion of individuals with triphasic (P0-present) EODs and blue indicates proportion of individuals with biphasic (P0-absent) EODs. Bongolo Falls is indicated in the map of Southern populations, and a grey region bounded by a dotted line indicates the small streams and creeks that drain into the Louétsi river above the Bongolo Falls.

The first goal of this study was to therefore explicitly test the hypothesis that genetic drift is responsible for the biogeographic distribution of EOD signals in *P. kingsleyae*. If this were the case, we would expect signal divergence to increase linearly with genetic divergence (Wilkins et al. 2013). Alternatively, if adaptive

forces were the main determinants of signal diversity, we would expect no correlation between signal divergence and genetic divergence.

A prominent alternative hypothesis for the distribution of signals is that mormyrid species may mate assortatively on

Table 1. Summary of locations and years of population samples, and analyses performed on each population.

Population	Year	Latitude	Longitude	<i>N</i>	Genotype	EOD	EO	Behavior
Apassa	2009	2°12'42" S	11°27'50" E	28	X	X	X (4)	
Bale	2001	0°31'9" S	12°47'58" E	8	X	X		
Bambomo	2009	2°9'49" S	11°27'42" E	106	X	X	X (7)	X (17)*
Bambomo	1999	2°9'49" S	11°27'42" E	27	X	X		
Bavavela	2009	2°14'33" S	11°33'22" E	24	X	X		
Bikagala	2009	2°11'43" S	11°33'40" E	28	X	X		
Mouvanga	2009	2°19'23" S	11°41'18" E	32	X	X	X (6)	X (10)*
Mouvanga	1999	2°19'23" S	11°41'18" E	19	X	X		
Nyamé-Pendé	2001	0°30'16" S	12°47'48" E	9	X	X		
Okano	2001	0°48'57" S	11°39'1" E	37	X	X		
Songou	2009	2°16'42" S	11°36'41" E	9	X	X	X (4)	

*Additional individuals were sampled from these populations to perform these analyses. Numbers in parentheses indicate sample sizes; if no sample size is reported, the entire *N* reported was analyzed. Column EO indicates population used in histological analysis of electric organs (see methods). Column Behavior indicates population used in EOD discrimination task (see methods). Table S1 contains the full metadata for all specimens, including Cornell Museum of Vertebrates accession numbers for vouchers.

the basis of EOD differences (Arnegard et al. 2005; Feulner et al. 2009a,b; Arnegard et al. 2010b), which would imply that EOD divergence is under sexual selection. For this to be a valid explanation for the present distribution of signal types within *P. kingsleyae*, it would require that *P. kingsleyae* have the perceptual ability to discriminate EOD differences. We tested this alternative hypothesis directly, using a dishabituation behavioral assay to determine whether *P. kingsleyae* from a sympatric location are able to discriminate between EOD types. We also assessed this hypothesis indirectly by examining patterns of genotypic differentiation and electric organ morphology to determine whether there was any evidence of assortative mating among *P. kingsleyae* EOD types.

Materials and Methods

FIELD COLLECTIONS

Collections of *Paramormyrops kingsleyae* specimens were made during field trips to Gabon, West Central Africa in 1999, 2001, and 2009 from nine locations summarized in Table 1 and Figure 2. A catalog of individual specimens used in this analysis is provided as Table S1, which provides Cornell Museum of Vertebrates accession numbers for all specimens used in this paper, along with associated metadata. We sampled two locations, Mouvanga and Bambomo Creeks, twice: once in 1999–2001 and again in 2009. Samples collected in different years at these locations were kept separate for all analyses, resulting in a total of 11 sampled populations.

Fish were collected using a variety of methods, including fish traps baited with worms, hand nets combined with electric fish detectors, and by light rotenone applications. Fol-

lowing any application of rotenone, we immediately transferred the fish to fresh, aerated water, where they recovered completely.

We georeferenced sampling locations and calculated pairwise geographic distances between all study populations using digitized topographic maps, which were superimposed over satellite imagery provided in Google Earth software (version 6.0.1; Google, Inc., Mountain View, CA). For each pair of populations, the distance between any two populations was assumed to be the shortest river path between the populations and was calculated by tracing currently mapped rivers between these populations. We note that additional, shorter paths connecting populations could potentially be created by seasonal flooding events (see Arnegard et al. 2006).

EOD RECORDINGS

Paramormyrops kingsleyae exhibit a sex difference in their signals during the rainy season breeding period, with sexually mature males showing a two- to threefold elongation of their EODs compared to female or nonbreeding males. Given the underrepresentation of breeding male recordings in our collections, the current analysis of electric signal variation is restricted to the species-typical female-like EODs exhibited by adult females and males outside the breeding season.

EODs of each specimen were originally recorded within 24 h of capture in 1- to 5-liter plastic boxes filled with water from the collection site. Signals were recorded with bipolar chloride-coated silver wire electrodes and amplified (bandwidth = 0.0001–50 kHz) with a differential bioamplifier (CWE, Inc., Ardmore, PA), and digitized at a 100 kHz–1 MHz sampling rate, with head-positivity plotted upward using a Daqbook or

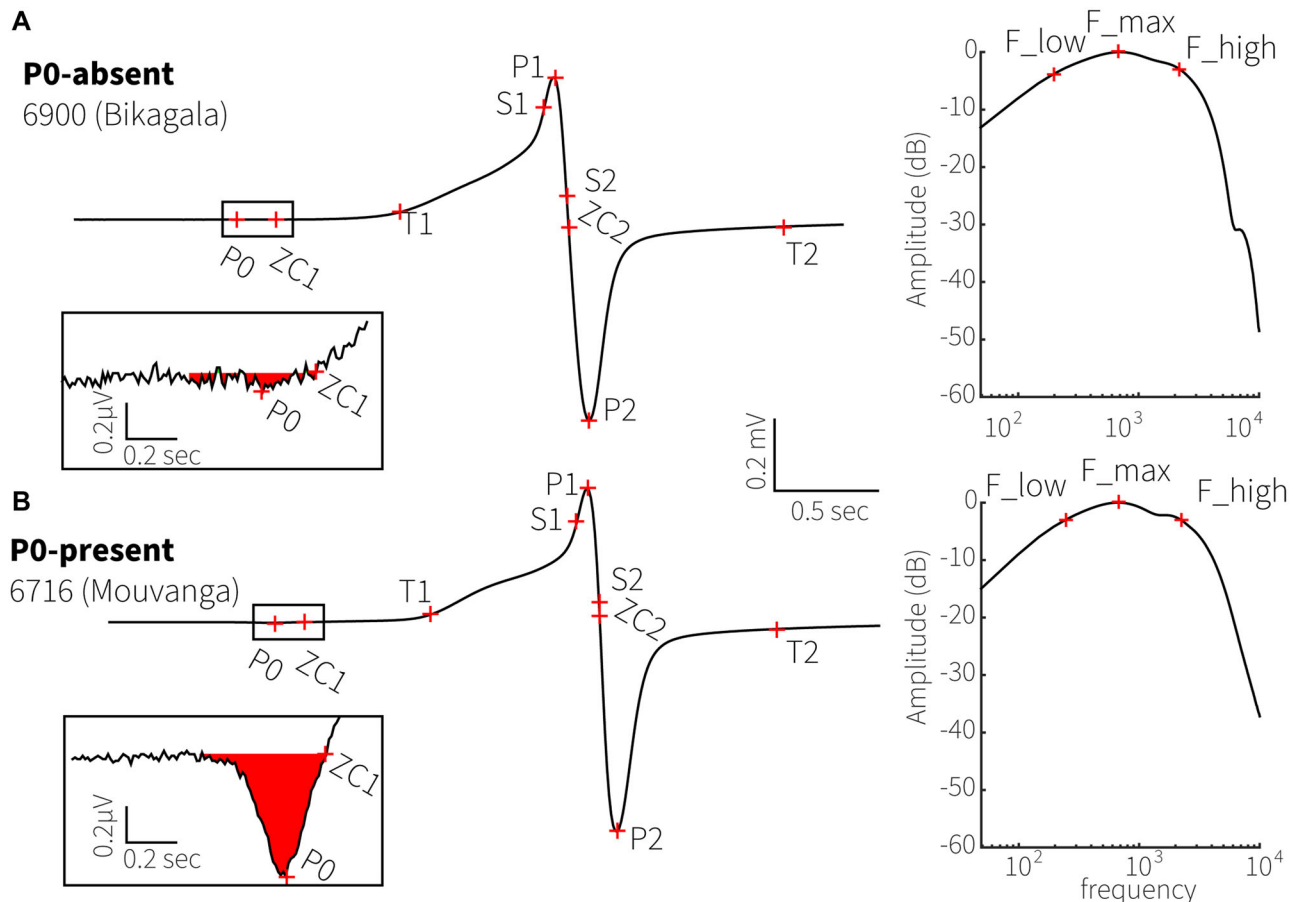


Figure 3. Example EOD Waveforms and Landmarks Measured. EOD landmarks and measurements used in the PCA were identical to those described in Gallant et al. (2011) and are listed for the reader's convenience in Table S2. We indicate two example EOD waveforms: biphasic (P0-absent) EOD. A. and triphasic (P0-present) EOD. B. to illustrate the locations of these landmarks on EOD waveforms and their associated power spectra. Voltages shown are relative to a normalized P1-P2 voltage set equal to 1.0 volts, as described in methods.

WaveBook (IOTECH, Cleveland, OH), or a USB-powered A-D Converter (National Instruments, Austin, TX). All EOD recordings were made at a vertical resolution of 16 bits per sample. After recording their EODs, we euthanized individual specimens with an overdose of MS-222. We removed one or more paired fins from specimens and preserved these tissues in 95% ethanol. Each specimen was given a unique specimen identification tag, and fixed free-floating in 10% formalin (phosphate-buffered; pH 7.2) for at least 2 weeks. Specimens were then transferred to 70% ethanol and deposited in the Cornell University Museum of Vertebrates. All methods conform to protocols approved by Cornell University's Center for Research Animal Resources and Education.

ANALYSIS OF ELECTRIC SIGNAL VARIATION

Following the methods described by Gallant et al. (2011) and Arnegard and Hopkins (2003), we made 21 measurements from each recorded EOD waveform using a custom written program

in MATLAB (Mathworks, Inc., Natick, MA). For each waveform, we calculated amplitudes, times, and slopes at nine landmarks defined by peaks, zero crossings, first derivative peaks, and threshold crossings (Fig. 3; Table S2). In addition, we calculated a power spectrum for each EOD waveform using the MATLAB *Fast Fourier Transform* function to determine the peak frequency and frequency values with power 3 dB below the peak frequency. We quantified patterns of EOD variation among all *P. kingsleyae* individuals by performing a principal components analysis (PCA) on the set of all 21 measurements using the function *princomp* in R version 3.4.3 (R Core Team 2017). The measurements were normalized with the function *scale*. Electric signal distance between populations was calculated as the Euclidean distance between group centroids for the first two principal components using the R package *vegan* (Oksanen et al. 2018). To test the null hypothesis of no differentiation between EOD waveforms from different populations, we also computed the Euclidean distance between all pairs of individuals and performed a permutational

MANOVA (PERMANOVA) directly on this pairwise signal distance matrix using 1000 permutations to determine probability values, using the *adonis* function from *vegan*.

Landmark-based signal processing methods as described above focus only on preselected signal features that do not encompass a comparison of the entire waveforms and may thus be discriminatory. Therefore, we complemented our PCA analysis by performing cross-correlation analysis of all 327 waveforms, as described in Carlson et al. (2011) and performed in Picq et al. (2016). EOD waveforms that were sampled at 150, 200, or 250 kHz ($n = 57$) were first down-sampled to 100 kHz using the *downsample* and *resample* functions in MATLAB. We then used the maximum of the absolute values of the cross-correlation function as a measure of pairwise waveform similarity, resulting in a matrix of pairwise similarities ranging from 0 (dissimilar waveforms) to 1 (identical waveforms). Multidimensional scaling (MDS) was then applied to this cross-correlation matrix using the “*mdscale*” function in Matlab with Kruskal’s normalized stress 1 criterion (Kruskal and Wish 1978). Electric signal distance between populations was calculated as the Euclidean distance between group centroids in the MDS space using *vegan*. Electric signal distances between all pairs of individuals were also estimated in the same way. As for the PCA, a PERMANOVA analysis was also performed on the individual pairwise signal matrix to test the null hypothesis of no differentiation between EOD waveforms from different localities.

Recording temperature is known to affect EOD duration (Kramer and Westby 1985). Gallant et al. (2011) showed that applying a Q10 temperature correction on 491 *P.kingsleyae* EODs recorded across Gabon at temperatures ranging from 21 to 26.7°C did not result in significantly different PCA scores of signal variation. As the range of recording temperatures in the current study is narrower (20.5–25.1°C), we considered the effect of temperature on our PCA configuration of signal variation as negligible. Nevertheless, we still investigated the effect of temperature on signal variation quantified through multidimensional scaling of signal cross correlations. The full methodology and results of this analysis are included in the Supporting Information File. We concluded from this analysis that recording temperature was not a significant source of variation in our study, and therefore present our analysis on the full dataset without temperature correction for both PCA-derived and MDS-derived signal analyses.

MICROSATELLITE GENOTYPING

We extracted DNA from the ethanol-preserved fin clips using DNeasy Tissue Kits (Qiagen, Inc., Hilden, Germany) for the 1998–1999 samples, or AgenCourt DNAadvance kits (Beckman-Coulter, Inc., Brea, CA) for the 2009 samples. The 1998–1999 DNA samples originally genotyped by Arnegard et al. (2005)

were re-genotyped for this study so that genotypes could be scored using identical criteria. We amplified DNA at each of five microsatellite loci (NBB001–NBB005) originally identified by Arnegard et al. (2005) using the Qiagen Type-It multiplex PCR System (Qiagen, Inc., Hilden, Germany). Reaction volumes were 15 µL, consisting of 1 µL template DNA, 7.5 µL Type-it Multiplex Master Mix (containing HotStarTaq Plus DNA Polymerase and PCR buffer with 6 µM MgCl₂), and 2 pmol of each primer (5′ primers labeled with fluorescent dyes FAM, HEX, or NED (Applied Biosystems, Foster City, CA)). Thermal cycling (under mineral oil) was 5 min at 95°C (initial activation) followed by 28 cycles of 95°C for 30 s, 60°C for 90 s, and 72°C for 30 s. Each individual reaction (containing PCR products for all 5 loci) was resolved by electrophoresis on an ABI 3100 automated sequencer (Applied Biosystems, Foster City, CA). Under these thermal conditions, reactions for locus NBB004 failed for the Bambomo, Nyamé-Pendé, and Balé Creek populations. For these populations, an additional PCR reaction was successfully performed as above using primers only for the NBB004 locus, with thermal cycling (under mineral oil) for 5 min at 95°C (initial activation) followed by 35 cycles of 95°C for 30 s, 50°C for 90 s, and 72°C for 30 s. Following genotyping, individual fragment lengths were analyzed and binned according to size by visual inspection, using Genemapper 4.1 software (Applied Biosystems, Foster City, CA).

GENOTYPING DATA ANALYSIS

For each microsatellite locus (NBB001–NBB005), we examined possible deviations from expected Hardy-Weinberg equilibrium within populations using the two-tailed exact test (Weir 1990) as implemented by GENEPOP version 4.1 (Rousset 2008). Next, we also performed exact tests of linkage disequilibrium between all pairs of loci (within and between populations) in GENEPOP to test the independent assortment of loci. Statistical significance in both sets of tests was evaluated using Markov chain methods (10,000 dememorization steps; 1000 batches; 5000 iterations per batch). We additionally calculated observed and expected heterozygosity under Hardy-Weinberg equilibrium for each locus in every population using the software Arlequin version 3.5 (Excoffier and Lischer 2010). The significance of each was evaluated at both the $P = 0.05$ α probability threshold and the Bonferroni corrected threshold of $P = 0.001$.

POPULATION STRUCTURE

We quantified genetic differentiation between populations using F_{st} (Weir and Cockerham 1984) with Arlequin version 3.5 (Excoffier and Lischer 2010). This standardized measure of population genetic structure is equivalent to the variance of allele frequencies between populations (i.e., subpopulations, or s) divided by the variance of allele frequencies in the total population

consisting of both subpopulations combined (t). Using the same software, we evaluated the statistical significance of the F_{st} estimates (at both the uncorrected and Bonferroni-corrected thresholds) by permuting genotypes between populations 50,000 times.

Because only five microsatellite loci were scored, we performed power analysis simulations using POWSIM version 4.1 (Ryman and Palm 2006) to determine if our sample sizes, number of microsatellite loci, and allele diversity were sufficient to detect genetic differentiation (see full methodology and results in Supporting Information File). These analyses showed that our sample sizes and specific genetic markers were adequate for detecting levels of genetic differentiation as low as $F_{st} = 0.003$ with a high probability (>0.9) (Fig. A2 in Supporting Information File). We can therefore consider our genetic markers as sufficiently powerful for the purpose of this study, namely, to give a reliable estimate of genetic differentiation between *P. kingsleyae* allopatric populations.

ISOLATION BY DISTANCE

Genetic isolation by distance (IBD) was explored following Rousset (1997), that is, using a linear regression between genetic distance ($F_{st}/(1 - F_{st})$) versus geographic distance for all pairs of populations. The correlation was tested with a Mantel test (Mantel 1967), which accounts for nonindependence of points in a distance matrix, using 1000 permutations in the R package *vegan*. The IBD analyses were performed on the whole dataset and repeated on Southern populations separated by short distances (<120 km) where sampling was high (including Bambomo, Apassa, Bikagala, Bavavela, Songou, and Mouvanga; Fig. 2 lower panel). When considering the entire dataset, it is important to note that populations included in this study represent two distant and separate zones along the species' otherwise continuous distribution across Gabon (Gallant et al. 2011). To control for this gap distribution in our sampling efforts and for the associated potential confounding effect of hierarchical structure (i.e. of regional-level effects between discontinuous Southern and Northern populations) on relationships between distance matrices, we ran a partial Mantel test, including a model matrix of regional membership that identified which populations were included in each pairwise comparison (e.g., North-North, South-South, South-North; Smouse et al. 1986; Meirmans 2012). When only considering the subset of Southern populations, we additionally tested for the effect of Bongolo Falls as a barrier to gene flow between upstream (Bavavela and Bikagala) and downstream (Bambomo, Apassa, Songou, and Mouvanga) populations by running a Mantel test between genetic distance and a model matrix coding for populations separated (1) or not separated (0) by this putative barrier.

COMPARISON OF SIGNAL AND GENETIC DIVERGENCE

To investigate whether electric signals evolve at a rate consistent with neutral evolution, we tested the correlation between signal and genetic distance for all pairs of populations using Mantel tests. This was performed for the whole dataset with a partial Mantel test (accounting for potential regional-level effects between discontinuous Northern and Southern populations) and repeated on Southern populations. We also tested whether signal differences between populations could be the product of IBD by testing the correlation between signal and geographic distances, again using a partial Mantel test for the whole dataset and a Mantel test for the Southern populations. The potential effect of Bongolo Falls in driving signal divergence in Southern populations was tested using the same rationale as for the genetic data: a Mantel test was performed between signal distance and a model matrix coding for populations separated or not by the falls. All analyses were performed on both PCA- and MDS-derived signal distances.

EVALUATING DISCRIMINATION ABILITY: BEHAVIORAL PLAYBACK EXPERIMENTS

We performed two sets of electrical playback experiments to assess the ability of *P. kingsleyae* to discriminate P0-present and P0-absent waveforms. As for previous analyses, these experiments were restricted to adult specimens exhibiting nonbreeding EODs. For all experiments, we assessed behavioral discrimination of EOD waveforms using a dishabituation paradigm described in detail by Carlson et al. (2011). Specimens were placed in a rectangular PVC enclosure containing both chlorided silver wire stimulus electrodes (Ag/AgCl) and Ag/AgCl recording electrodes, with the two pairs of electrodes oriented orthogonally with respect to one another. We delivered stimulus trains consisting of 10 bursts of 10 EOD pulses each, with an intraburst interval of 30 ms, and interburst interval of 10 s, with a peak-to-peak intensity of 145 mV/cm. We constructed negative control trains in which all 10 bursts of EOD pulses were identical. We also constructed positive controls known to exhibit reliable responses, using eight bursts of identical EODs (background), with the ninth burst consisting of a novel waveform (a 90-degree phase-shifted EOD used during the first eight bursts), followed by a 10th burst of the original background waveform. Positive control novel EOD stimuli were constructed by phase shifting the background EOD waveform by 90°, which is maximally divergent in the time domain while preserving the frequency spectrum. Phase shifting of EODs was accomplished by performing a fast Fourier Transform (FFT) on the EOD waveform, followed by adding 90° to the phase angle at each positive frequency, and subtracting 90° from each the phase angle at each negative frequency. This was followed, in turn, by inverse

FFT, which yielded a reconstructed EOD characterized by an altered (i.e., 90° shifted) phase spectrum with an unaltered power spectrum (Heiligenberg and Altes 1978; Hopkins and Bass 1981; Carlson and Arnegard 2011). *Paramormyrops kingsleyae* have been shown to respond to this specific phase-shifted experimental stimulus by increasing their electrical discharge rate (Carlson et al. 2011). This trial was thus included in our experiments to demonstrate that the fish are capable of exhibiting electrical responses when they are presented with electrical stimuli that they can discriminate. The experimental stimuli consisted of presenting eight bursts of the same *P. kingsleyae* EOD with the ninth burst consisting of a different *P. kingsleyae* EOD. We computed the EOD rate of the subject fish by converting the EOD times of occurrence into a series of a delta functions, and convolving these a 300-ms Gaussian waveform (Carlson and Hopkins 2004). The specimen's response was then recorded as the maximum EOD rate within a 2-s window following each burst of EODs (Carlson and Arnegard 2011). The magnitude of the specimen's response declined over repeated presentation of the background bursts (habituation). Therefore, we measured each specimen's "novelty response" as the change in EOD discharge rate (dishabituation) following the ninth (novel) burst, relative to the EOD discharge rate following the eighth presentation of the background burst. Statistical significance in signal discrimination was assessed using Dunnett's multiple comparison test with control (Dunnett 1955), whereby we compared novelty responses toward each experimental stimulus (and toward the positive control) with the negative control. Significantly different novelty responses to experimental stimuli versus negative control were considered evidence for discrimination between background and novel waveforms.

Some trials included "hybrid" waveforms generated artificially from two normalized natural EOD waveforms: one P0-absent waveform from Bambomo and one P0-present waveform from Mouvanga. We centered these waveforms using the midpoint between peaks P1 and P2 of the waveform, and generated hybrids of varying P0 character by taking different weighted averages of the two waveforms (Fig. S1).

One set of experiments was performed within 6 h of capture on P0-absent *P. kingsleyae* specimens from Bambomo creek ($n = 10$), where both signal types are known to co-occur, as well as on P0-present *P. kingsleyae* individuals from Mouvanga creek ($n = 10$), where only P0-present *P. kingsleyae* individuals occur but other *Paramormyrops* species are found. A second set of experiments was performed on additional P0-absent individuals from Bambomo Creek, which were captured as juveniles and transported to the Hopkins laboratory at Cornell University (Ithaca, NY), and reared to adulthood for further study ($n = 13$). These fish were housed in community tanks with other individuals captured from their home stream. In addition, the fish were

fed live blackworms daily, and they were maintained on a 12-h light/dark cycle, with water temperatures maintained between 25 and 27°C, pH between 6.5 and 7.0, and water conductivity between 200 and 400 μ S.

ELECTRIC ORGAN HISTOLOGY AND ANALYSIS

Serial sections of electric organs from selected individuals were made for light microscopy analysis following the methods described by Gallant et al. (2011). Briefly, electric organs were removed from fixed specimens, decalcified overnight, dehydrated in a graded alcohol series, and infiltrated in glycol methacrylate resin (JB-4 resin; Polysciences, Inc., Warrington, PA). Serial parasagittal sections, each 6- μ m thick, were cut from lateral to medial with a tungsten carbide microtome knife, mounted on glass slides, and stained with 0.5% Toluidine blue for 30 s. For each specimen, we reconstructed one of four columns of electrocytes from these serial sections. Because each column of electrocytes surrounds the spinal cord, we began our reconstruction at the lateral edge of the electric organ, and stopped the reconstruction when the spinal cord was clearly visible (approximately 234–648 μ m depending on the size of the individual). For each section, the number of stalk penetrations through each electrocyte was counted in, and averaged across 50–70 electrocytes per section. An electrocyte was scored with a penetration whenever a stalk was observed to pass through either one or both faces of the electrocyte. For our analysis, we considered each 6- μ m section to have an independent number of penetrations from all other sections to minimize the probability of underestimating the total number of penetrations.

Results

PATTERNS OF EOD VARIATION

We assessed variation in EOD signals using both PCA (Fig. 4) and multidimensional scaling of signal cross correlations (MDS; Fig. S2) on each of the 327 individual EODs in this study.

The first principal component of our PCA related strongly to duration and explained 43.71% of the variation between individuals, whereas the second factor related strongly to the magnitude of P0 and comprised 15.94% of the variation between electric signals. Factor loadings for these two principal components are summarized in Table S3. In our MDS analysis, the number of dimensions was set to $N = 2$, which resulted in a stress of 0.0526, considered to give a good ordination representation of the cross-correlation matrix with low probability of misinterpretation (Clarke 1993).

Both MDS and PCA identified significant variation among recording localities (PERMANOVA on PCA signal distances: $F_{10,326} = 58.62$, $P < 0.001$; PERMANOVA on MDS signal distances: $F_{10,326} = 45.44$, $P < 0.001$). Mantel tests showed a

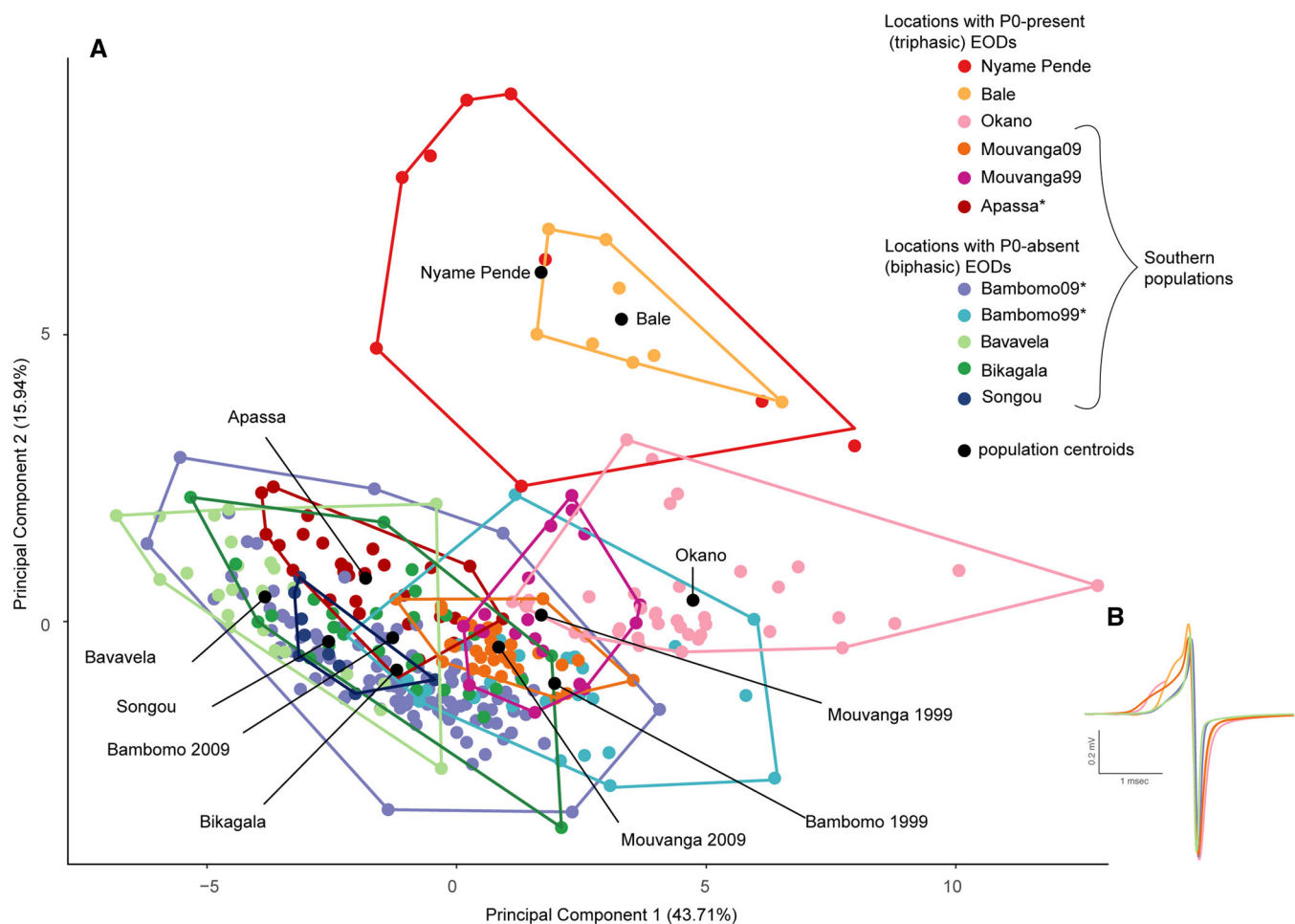


Figure 4. Principal component analysis of EOD waveform variation in 327 *P. kingsleyae* individuals from 9 populations in Gabon. **A.** Variation in waveforms was quantified using Principal Components Analysis (see Gallant et al. (2011) and text for further details). Variables related primarily to EOD duration loaded most strongly on principal component 1, whereas variables related to the magnitude of P0 loaded most strongly on principal component 2 (Table S3). Polygons enclose EOD waveforms from each recording locality. Polygon centroids are represented with black dots. Asterisks in the legend represent two populations with mixed signal types (Apassa is mostly composed of P0-present individuals, whereas Bambomo is mostly composed of P0-absent individuals). **B.** Overlay diagram of EOD waveforms from five different localities.

strong and significant correlation between signal distances estimated from PCA and MDS (Fig. S3, Mantel test, $R = 0.677$, $P = 0.001$), which is also evident from comparison of Figures 4 and S2. Interestingly, inter-population distances between Southern populations and Bale as well as between Southern populations and Nyamé-Pendé were larger with PCA-derived methods than with MDS-derived methods (Fig. S3). This is likely due to the fact that the PCA analysis included three variables pertaining to the frequency content of the EODs (peak frequency, low and high frequency with 3dB below the peak frequency) in addition to variables pertaining to the temporal content of the EODs. The cross-correlation analysis that the MDS was applied to, on the other hand, involves the progressive sliding of one waveform past the other, and thus indicates EOD similarity essentially in temporal content. Although a change in frequency content will be

associated with a change in temporal content, the MDS analysis likely attributed less weight to specific frequency content differences. Regarding the two populations that were sampled in 1999 and again in 2009, we note that the overlap between population polygons corresponding to different sampling years is more extensive for Mouvanga than for Bambomo in both PCA and MDS, suggesting more extensive EOD divergence in Bambomo than in Mouvanga within a period of 10 years.

PATTERN OF GENETIC VARIATION

We were able to amplify fragments without any failed reactions (i.e., possible null homozygotes) for all 327 individuals genotyped in this study. Total number of alleles detected at each of five loci over all populations ranged from 2 to 21 (Table 2). For each population, locus-specific expected heterozygosities ranged

Table 2. Summary of loci attributes.

Population	NBB01				NBB02				NBB03				NBB04				NBB05			
	<i>k</i>	<i>H_o</i>	<i>H_e</i>	<i>P</i>	<i>k</i>	<i>H_o</i>	<i>H_e</i>	<i>P</i>	<i>k</i>	<i>H_o</i>	<i>H_e</i>	<i>P</i>	<i>k</i>	<i>H_o</i>	<i>H_e</i>	<i>P</i>	<i>k</i>	<i>H_o</i>	<i>H_e</i>	<i>P</i>
Apassa	6	0.71	0.67	0.955	4	0.79	0.71	0.393	6	0.75	0.68	0.212	4	0.64	0.75	0.276	17	0.75	0.92	0.184
Bale	4	0.50	0.53	0.655	8	0.75	0.89	0.432	7	0.75	0.85	0.762	3	0.50	0.49	0.283	9	0.75	0.92	0.241
Bambomo 09	6	0.37	0.35	0.953	9	0.82	0.73	0.135	7	0.68	0.68	0.259	10	0.38	0.62	0.000	21	0.72	0.79	0.183
Bavavela	6	0.67	0.60	0.693	4	0.58	0.59	0.184	6	0.83	0.81	0.546	4	0.83	0.73	0.059	6	0.71	0.71	0.584
Bambomo 99	4	0.41	0.38	0.537	7	0.67	0.77	0.251	6	0.44	0.64	0.006	8	0.22	0.69	0.000	13	0.78	0.81	0.455
Bikagala	5	0.68	0.64	0.485	4	0.50	0.50	0.04	4	0.50	0.59	0.437	4	0.34	0.33	0.373	4	0.57	0.54	0.053
Mouvanga 09	2	0.09	0.09	1.000	3	0.16	0.15	1.000	4	0.25	0.23	1.000	6	0.44	0.43	0.445	3	0.44	0.51	0.576
Mouvanga 99	2	0.05	0.05	1.000	3	0.16	0.15	1.000	5	0.26	0.41	0.023	6	0.37	0.48	0.228	4	0.63	0.56	0.644
Nyamé-Pendé	2	0.11	0.11	1.000	10	0.67	0.91	0.046	7	1.00	0.81	0.691	2	0.00	0.47	0.004	10	0.89	0.88	0.993
Okano	7	0.53	0.55	0.271	2	0.24	0.25	0.569	7	0.42	0.37	1.000	8	0.45	0.43	0.489	16	0.82	0.90	0.197
Songou	3	0.22	0.57	0.014	2	0.11	0.11	1.000	4	0.33	0.65	0.12	2	0.22	0.52	0.172	2	0.22	0.21	1.000

For each locus and population, the number of alleles (*k*) at each locus, the observed (*H_o*), and expected heterozygosity (*H_e*) are reported. Probabilities of deviation from Hardy-Weinberg equilibrium (HWE) are reported as *P*.

from 0.05 to 0.92 (Table 2). Exact tests produced no evidence of linked loci across all populations ($P > 0.17$). Of 55 unique locus-by-population combinations, only a few cases exhibited evidence of a deviation of observed heterozygosity from the expectation under Hardy-Weinberg equilibrium (at uncorrected $P = 0.05$, see Table 2). After Bonferroni adjustment for multiple comparisons, only the NBB004 samples for Bambomo 1999, Bambomo 2009, and Nyamé-Pendé remained significant.

The allele frequency histograms at each of five loci for each of the 327 individuals in the full dataset are shown in Figure 5. For the two populations that were sampled in 1999 and again in 2009 (Mouvanga and Bambomo), allele frequencies appeared to change little over this 10-year period between sampling. Between populations, there are apparent differences between the distribution of alleles; of particular note are the presence of several alleles (NBB002, ~250 bp; NBB005 several alleles between 300 and 400 bp), which are present in Bambomo and Apassa creeks but absent in other populations.

F_{st} values of all pairwise comparisons between the *P. kinglyae* populations surveyed are shown in Figure 6. All populations were significantly differentiated from one another at the $P = 0.05$ and the Bonferroni-corrected threshold ($P = 0.001$), with the exceptional pairwise comparisons of Bambomo Creek 1999 versus 2009, Mouvanga Creek 1999 versus 2009, and Nyamé-Pendé versus Bale Creek. Among all pairs of populations, the magnitude of genetic differentiation between populations varied from $F_{st} = 0.055$ to 0.65. We note that Mouvanga was highly differentiated from all other populations, with F_{st} values ranging from 0.377 (Mouvanga99-Nyamé-Pendé) to 0.65 (Mouvanga09-Songou). Most importantly, the populations of Bambomo (predominantly biphasic individuals) and Apassa (predominantly triphasic individuals), which represent a potential hybrid zone, exhibit the lowest F_{st} values of our dataset ($F_{st} < 0.06$; Fig. 5), despite being phenotypically distinct with respect to P0-presence/absence.

ISOLATION BY DISTANCE

We investigated the relationship between genetic differentiation and geographic distance, measured as shortest river distance between populations (Fig. 7A). Among all population pairs in Gabon, we found no significant relationship between genetic and geographic distance (Mantel test: $R = -0.042$, $P = 0.568$), even after correcting for potential regional-level effects (partial Mantel test: $R = 0.099$, $P = 0.278$). These results were consistent across different genetic distance measures (Table S4) or when samples collected in different years in Bambomo and Mouvanga were treated as one instead of separate populations (Tables S5 and S6).

In contrast, among population pairs restricted to the South of Gabon, genetic distance was strongly related to geographic

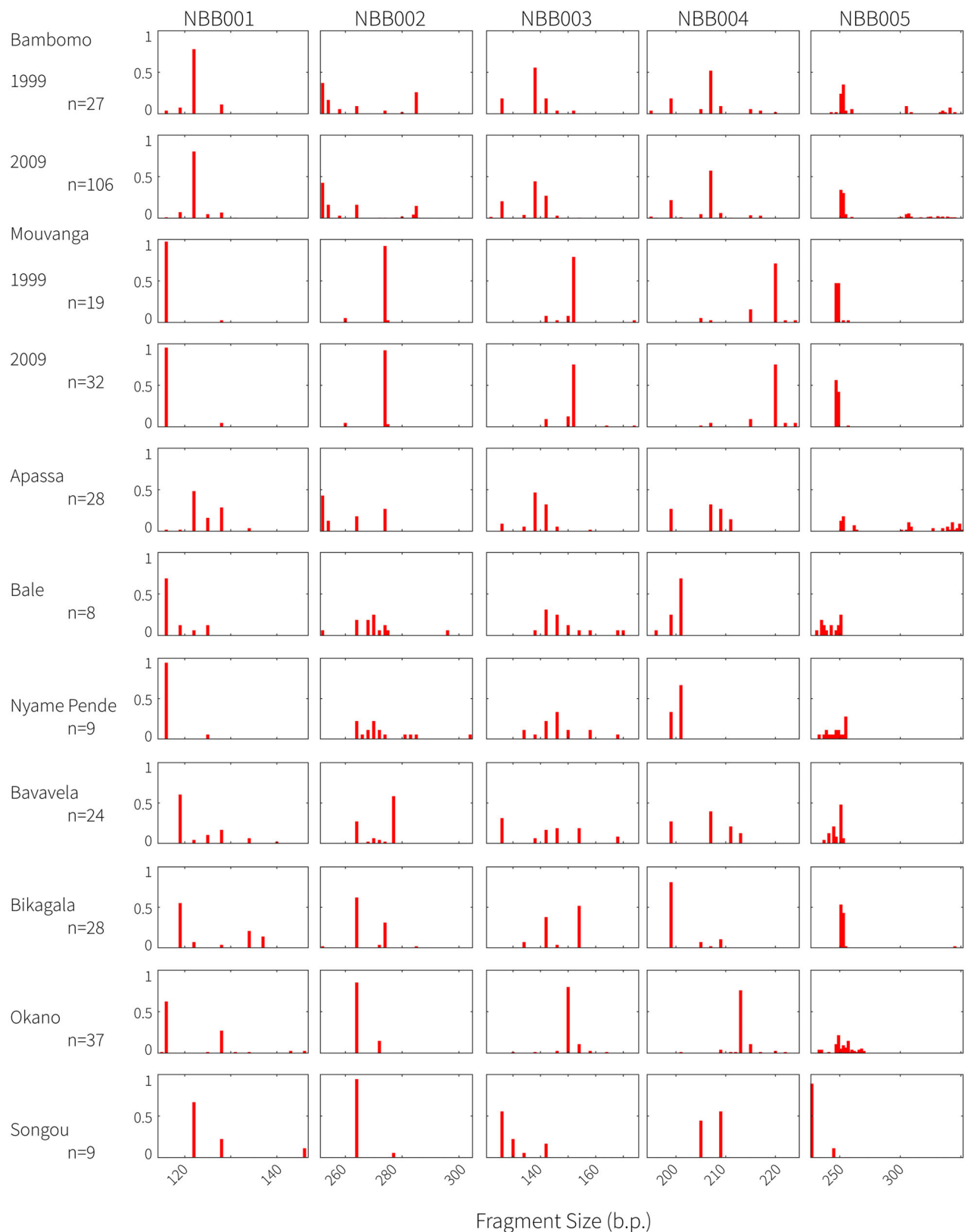


Figure 5. Allele frequency histograms for each population and microsatellite locus. Microsatellite loci were designated as NBB001-NBB005 by Arnegard et al. (2005). For each population, sample sizes are reported as number of individuals genotyped, so the plots include twice as many allele copies.

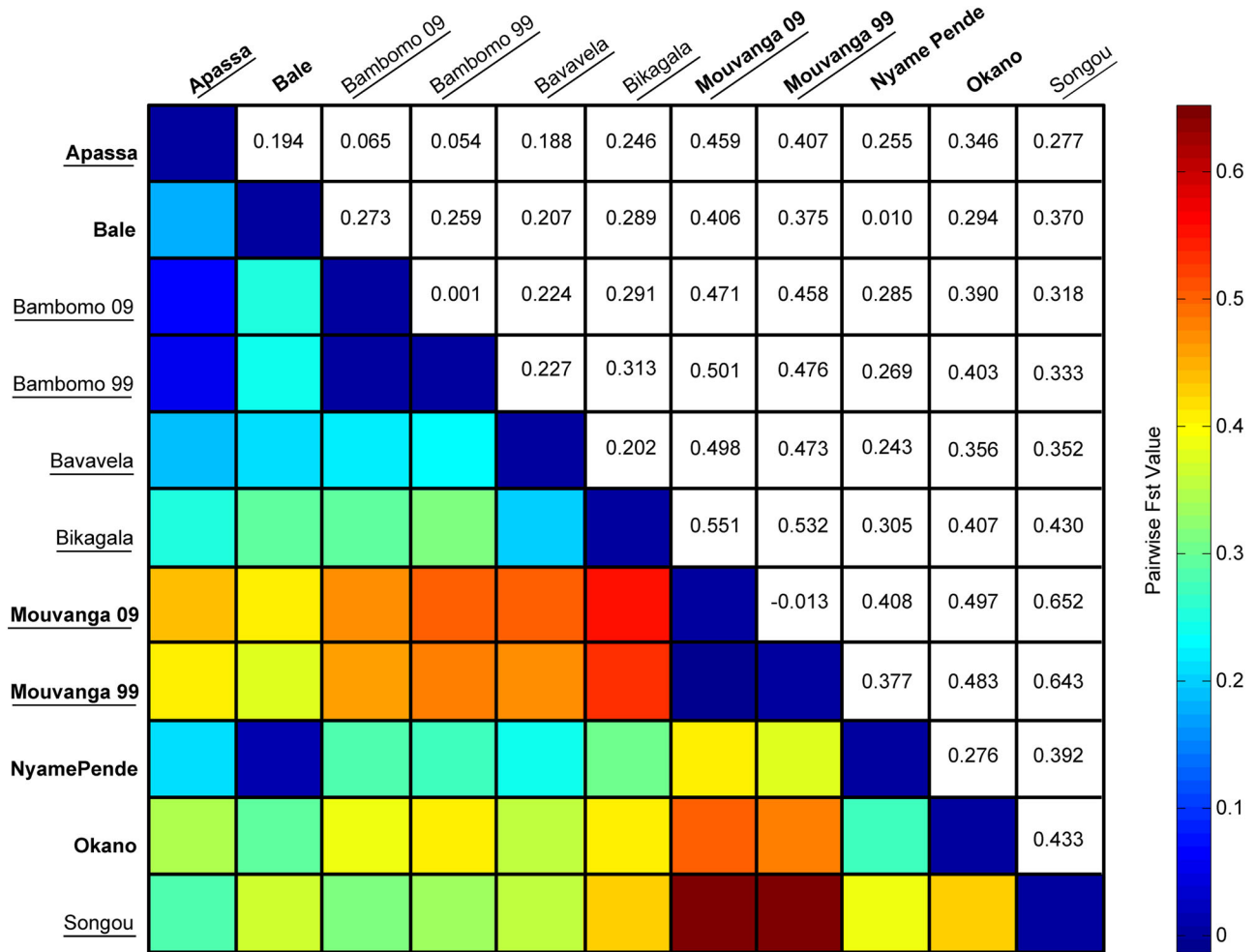


Figure 6. F_{ST} values for each pairwise comparison of populations. The lower half of the matrix codes F_{ST} value by color and the upper half of the matrix reports the individual F_{ST} value. All pairwise comparisons of F_{ST} values were significant at Bonferroni-corrected thresholds, with the exception of Mouvanga Creek (1999 vs. 2009), Bambomo Creek (1999 vs. 2009), and Balé Creek vs. Nyamé Pendé Creek. Underlined sites represent Southern populations. Localities in bold represent sites with triphasic (P0-present) EODs, whereas non-bolded localities indicate sites with biphasic (P0-absent) EODs.

distance (Fig. 7A; $R = 0.576$, $P = 0.02$), regardless of the genetic distance metric used (Table S4) or whether Bambomo and Mouvanga samples from different years were treated as independent populations or not (Tables S5 and S6). However, we found that genetic distances between populations separated by Bongolo falls were on average no greater than those between populations on the same side of the Falls (Fig. S4). This was confirmed by the lack of correlation between genetic distance and the Bongolo model matrix (Table 3).

CORRELATION BETWEEN SIGNAL, GENETIC, AND GEOGRAPHIC DISTANCES

We investigated the relationship among signal, genetic, and geographic distance (Fig. 7B–E) at two spatial scales (for correlation plots focusing only on Southern population, see Fig. S5A–

D). When considering the entire dataset, signal distances were not correlated with genetic distances (Figs. 7B and 7D; Table 3), even after correcting for potential regional-level effects (partial Mantel test: $R = 0.025$, $P = 0.438$). These results were consistent regardless of how signal or genetic distances were estimated (Tables 3 and S4) and of whether samples collected from different years in Bambomo and Mouvanga were considered as one instead of separate populations (Tables S5 and S6).

On the other hand, we found a strong and significant relationship between signal and geographic distance using PCA-derived signal distances (Fig. 7C; Table 3), even after correcting for potential regional-level effects (partial Mantel test: $R = 0.421$, $P = 0.03$) and after treating samples collected from different years as one instead of separate populations (Tables S5 and

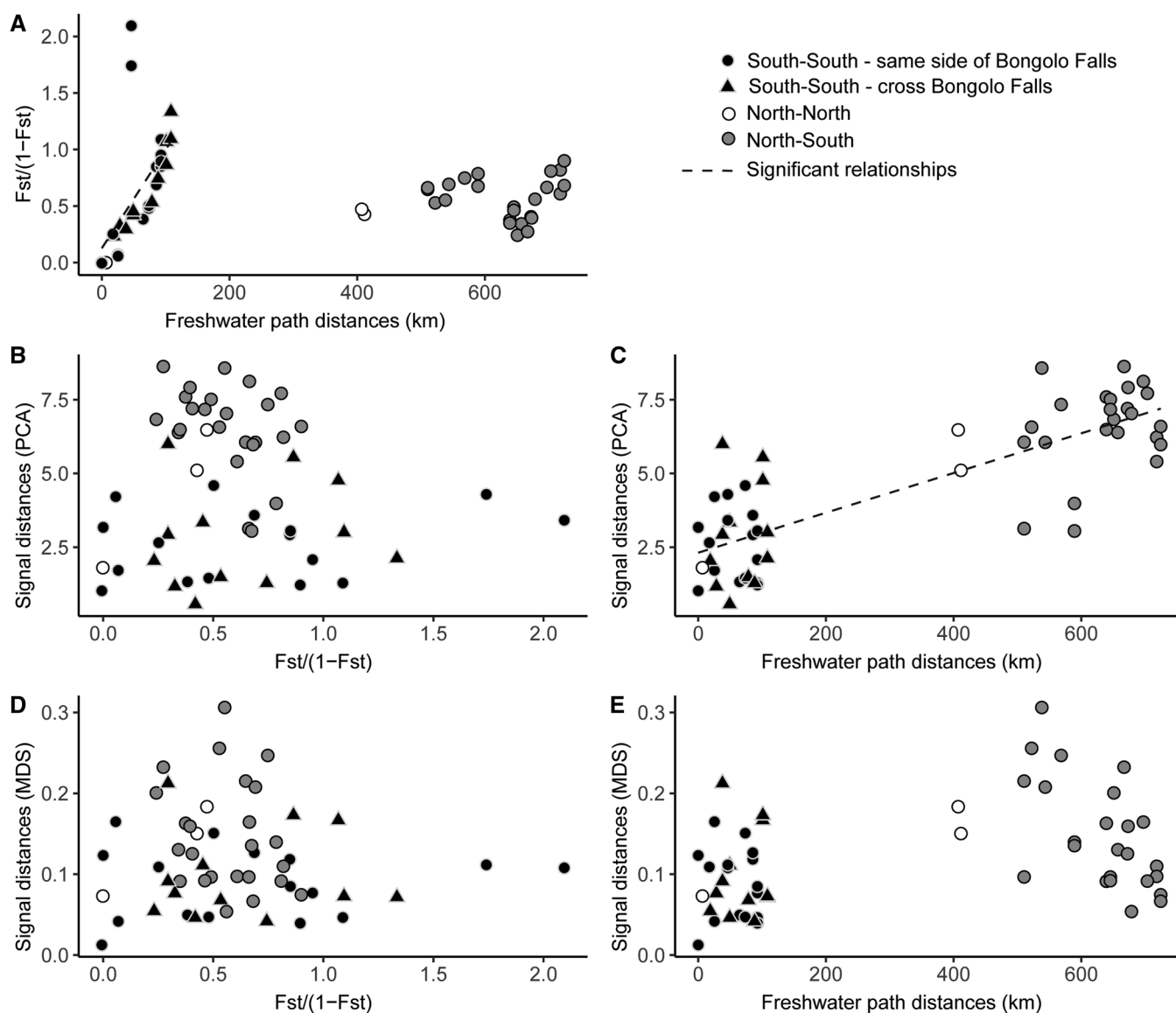


Figure 7. Correlation plots between genetic, signal and geographic distances for all populations ($n = 11$ populations, all symbols) and within Southern populations only ($n = 8$ populations, black symbols). A. Population pairwise genetic distance in relation to geographic distance. B. Correlation between PCA-derived signal and genetic distances. C. Correlation between PCA-derived signal and geographic distances. D. Correlation between MDS-derived signal and genetic distances. E. Correlation between MDS-derived signal and geographic distances. Significant relationships identified by Mantel tests are shown with a black dashed fit line (see Table 3).

S6). Using MDS-derived signal distances, the signal-geography correlation was weaker yet significant (Fig. 7E; Table 3); however, it disappeared when we controlled for regional-level effects (partial Mantel test: $R = -0.063$, $P = 0.603$). The same results were found regardless of how Bambomo and Mouvanga samples were grouped (Tables S5 and S6).

When considering only Southern populations, none of the variables tested (genetic distance, geographic distance, and Bongolo Falls model matrix) were found to correlate with signal distances (Fig. S5A–D; Tables 3, S4, S5, and S6).

***P. kingsleyae* CAN DETECT INTRASPECIFIC EOD WAVEFORM VARIATION**

We used a habituation-dishabituation paradigm (Carlson et al. 2011) to determine whether individual *P. kingsleyae* were capable of discriminating between sympatric and allopatric *P. kingsleyae* EOD waveforms that were either P0-absent or P0-present (Figs. 8A and 8B). We performed two sets of experiments to assess this: the first set of experiments was performed with single stimulus presentations and recordings of responses across 20 field-captured individuals (Figs. 8C and 8D), whereas the second

Table 3. Results of standard and partial Mantel tests between genetic distances ($F_{st}/(1 - F_{st})$), signal distances (obtained through PCA [left] and MDS [right]), geographic distances (km), and presence/absence of Bongolo Falls (Southern populations) at two spatial scales.

		PCA-derived signal distances		MDS-derived signal distances	
		Mantel's r	P -value	Mantel's r	P -value
Total range ($n = 11$ populations)	Correlation				
	Genetic - geographic	−0.042	0.568	−0.042	0.568
	Signal - genetic	−0.041	0.554	−0.051	0.559
	Signal - geographic	0.813	0.003	0.369	0.037
	Partial correlation				
	Genetic - geographic (regional clusters as covariate)	0.099	0.278	0.099	0.278
	Signal - genetic (regional clusters as covariate)	0.025	0.438	−0.023	0.502
	Signal - geographic (regional clusters as covariate)	0.421	0.030	−0.063	0.603
Southern populations ($n = 8$ populations)	Correlation				
	Genetic - geographic	0.576	0.021	0.576	0.021
	Genetic - Bongolo Falls	−0.042	0.550	−0.042	0.550
	Signal - genetic	0.181	0.175	0.062	0.303
	Signal - geographic	0.069	0.324	0.018	0.365
	Signal - Bongolo Falls	0.081	0.314	0.110	0.272

Significance at the 0.05 level or below is marked in bold.

was performed with five repetitions of randomly interleaved stimulus trains to laboratory-maintained individuals, which enabled us to average responses and thereby reduce variability (Fig. 8E–H; see Methods).

In the first field experiment (Fig. 8C), P0-absent *P. kingsleyae* from Bambomo creek showed behavioral evidence of discriminating phase-shifted EODs from P0-absent EODs ($P < 0.001$, as compared to negative control), but did not show evidence of discriminating between P0-absent EODs (background) and Mouvanga P0-present EODs or Bambomo P0-present EODs (novel). In the second field experiment (Fig. 8D), P0-present *P. kingsleyae* from Mouvanga creek showed greater dishabituation to phase-shifted EODs than test EODs, but this difference was not significant ($P = 0.1751$). There was no significant difference in response between Mouvanga P0-present EODs (background) and Bambomo P0-absent EODs or Bambomo P0-present EODs (novel). Taken together, these field experiments performed on *P. kingsleyae* from Bambomo Creek and Mouvanga creek did not support the hypothesis that *P. kingsleyae* are able to discriminate between the subtly different P0-absent and P0-present waveforms. We therefore performed laboratory experiments that were designed to be more sensitive at detecting small differences in response through averaging.

In the first laboratory experiment (Fig. 8E), we tested the hypothesis that *P. kingsleyae* from Bambomo Creek are able to discriminate allopatric P0-present EODs from sympatric P0-absent EODs. We were able to determine a statistically significant response to the positive control (novel = 90° phase-shifted EOD; $P < 0.001$). In addition, we detected statically significant changes in novelty responses to stimulus trains where both Mouvanga P0-present EODs were novel ($P < 0.01$), and where Bambomo P0-absent EODs were novel ($P < 0.001$). This evidence is consistent with the hypothesis that *P. kingsleyae* are able to discriminate between P0-present and P0-absent waveforms, but is confounded by the fact that allopatric populations, and even individuals, can differ slightly in EOD waveform (Gallant et al. 2011). Thus, there may be cues other than P0 that mediated this discrimination.

To rule out the possibility of individual EOD discrimination on the basis of characters *other than* P0-presence/absence, we tested *P. kingsleyae* captured in Bambomo Creek with individual sympatric EOD waveforms where P0 *was not* available as a cue (Fig. 8F). Although subjects responded significantly to positive controls (+90° and Mouvanga that have large P0-absent waveforms; $P < 0.001$), subjects did not exhibit a significant novelty response when presented background and novel stimuli that were all P0-absent ($P = 0.3478$), or when background and novel stimuli were all P0-present ($P = 0.3869$). In a third, corollary

laboratory experiment, we tested the hypothesis that *P. kingsleyae* from Bambomo Creek are able to discriminate between sympatrically occurring EOD variants (Fig. 8G) where P0-presence/absence was available as a cue. Subjects responded significantly to positive controls ($P < 0.001$), and to P0-present waveforms from Bambomo in a background of P0-absent waveforms from Bambomo ($P = 0.02$), although not to P0-present waveforms from Ivindo (which have very large P0s; $P = 0.7189$) from a background of P0-absent waveforms. Together, these three laboratory experiments support the hypothesis that *P. kingsleyae* can differentiate between sympatric EOD waveforms when P0 is available as a cue.

Finally, in the fourth laboratory experiment (Fig. 8H), we examined the ability of *P. kingsleyae* to discriminate between artificial EOD waveforms that had very small P0 peaks present (1:1 P0-absent/P0-present hybrid; see Methods), and natural EOD waveforms that had no P0-present. We determined a statistically significant response to positive controls ($P < 0.01$), and to EODs that represented 1:3 ratios of P0-absent to P0-present waveforms ($P < 0.01$), however, subjects did not elicit a statistically significant response to 3:1 ($P = 0.873$) or 1:1 ratio ($P = 1.0$) of P0-absent to P0-present EODs.

The results of the field and laboratory experiments are consistent with the hypothesis that *P. kingsleyae* can differentiate between P0-absent and P0-present waveforms.

MORPHOLOGICAL ANALYSIS OF ELECTRIC ORGANS IDENTIFIES ADDITIONAL SPECIMENS WITH MIXED ANATOMY IN APASSA CREEK

Gallant et al. (2011) showed that P0-absent EODs are produced by fish with *NPP* electrocyte anatomy (posterior innervation with non-penetrating stalks), whereas P0-present EODs were produced by fish with *Pa* electrocytes in the electric organ (anterior innervation with penetrating stalks). In addition, Gallant et al. (2011) presented evidence for electric organs with mixed morphology, wherein some electrocytes had penetrating stalks, whereas other electrocytes from the same organ had nonpenetrating stalks. We confirmed this observation with an additional analysis of 21 electric organs collected in 2009. Our present analysis supports the existence of these morphological types. In Figure 9, we show four examples of electric organs surveyed in Bambomo, Apassa, Mouvanga, and Songou Creeks, and summarize our analysis of all 21 individuals in Table 4. In Apassa and Mouvanga all individuals were of the *Pa* morphology type, with one exceptional individual in Apassa exhibiting mixed morphology (*NPP* + *Pa* morphology in the same individual). In Bambomo Creek and Songou creek, we detected individuals that had entirely *NPP* type morphology, with one exceptional individual in Bambomo creek that had entirely *Pa* type morphology.

Discussion

This study combined signal, genetic, and morphological analyses with behavioral experiments to investigate the evolutionary forces driving the intraspecific polymorphism of biphasic and triphasic signals in *P. kingsleyae*. This variation provides a rare window into the early stages of divergence in a key character that consistently varies between species in the *Paramormyrops* radiation and can thus inform us on the evolutionary processes that triggered this explosive signal diversification.

A key prediction of the drift hypothesis is that signal variation should be highly correlated with variation in neutral genetic markers. We found that signal distances were not related to genetic distances at any scale of our study, implying that although genetic drift is observable between populations in our dataset, it is insufficient to explain the evolution of diversity in the waveforms of this species. On the other hand, our behavioral experiments demonstrated that *P. kingsleyae* possess the perceptual ability to discriminate between biphasic and triphasic EODs, suggesting that EOD divergence in this species has the potential to be used as a cue for assortative mating with respect to signal type. This finding is consistent with previous studies that reported species-specific signal preferences among *Paramormyrops* species (Arnegard et al. 2006; in which *Brienomyrus* sp. CAB refers to *P. kingsleyae*) as well as among the other major mormyrid radiation of *Campylomormyrus* species (Feulner et al. 2009a,b; Nagel et al. 2018a,b).

THE RELATIONSHIP AMONG GENETIC, GEOGRAPHIC, AND PHENOTYPIC DISTANCE

Because the biogeographic distributions of tropical freshwater fishes are mainly constrained by landscape and ecological features such as basin geomorphology and river capture dynamics, differentiation patterns in these taxa at large spatial scales are often purely the result of vicariance and drift (Albert and Reis 2011). It is therefore unsurprising that we found a significant relationship between genetic and geographic distances in Southern Gabon, as would be predicted with an IBD model (Wright 1943). Despite this, we draw the reader's attention to a few key observations regarding the patterns of genetic differentiation between populations. First, we demonstrate that the presence of Bongolo Falls did not explain any of the genetic structure in this region (Tables 2, S3, S4, and S5), implying that populations separated by this 15-m waterfall are not necessarily genetically more diverged than populations not separated by it, which may be caused by occasional downstream migration. Second, we were unable to find a significant relationship between genetic and geographic distance that extended over large geographic scales. The observed pattern in *P. kingsleyae*'s genetic structure in our dataset across Gabon is

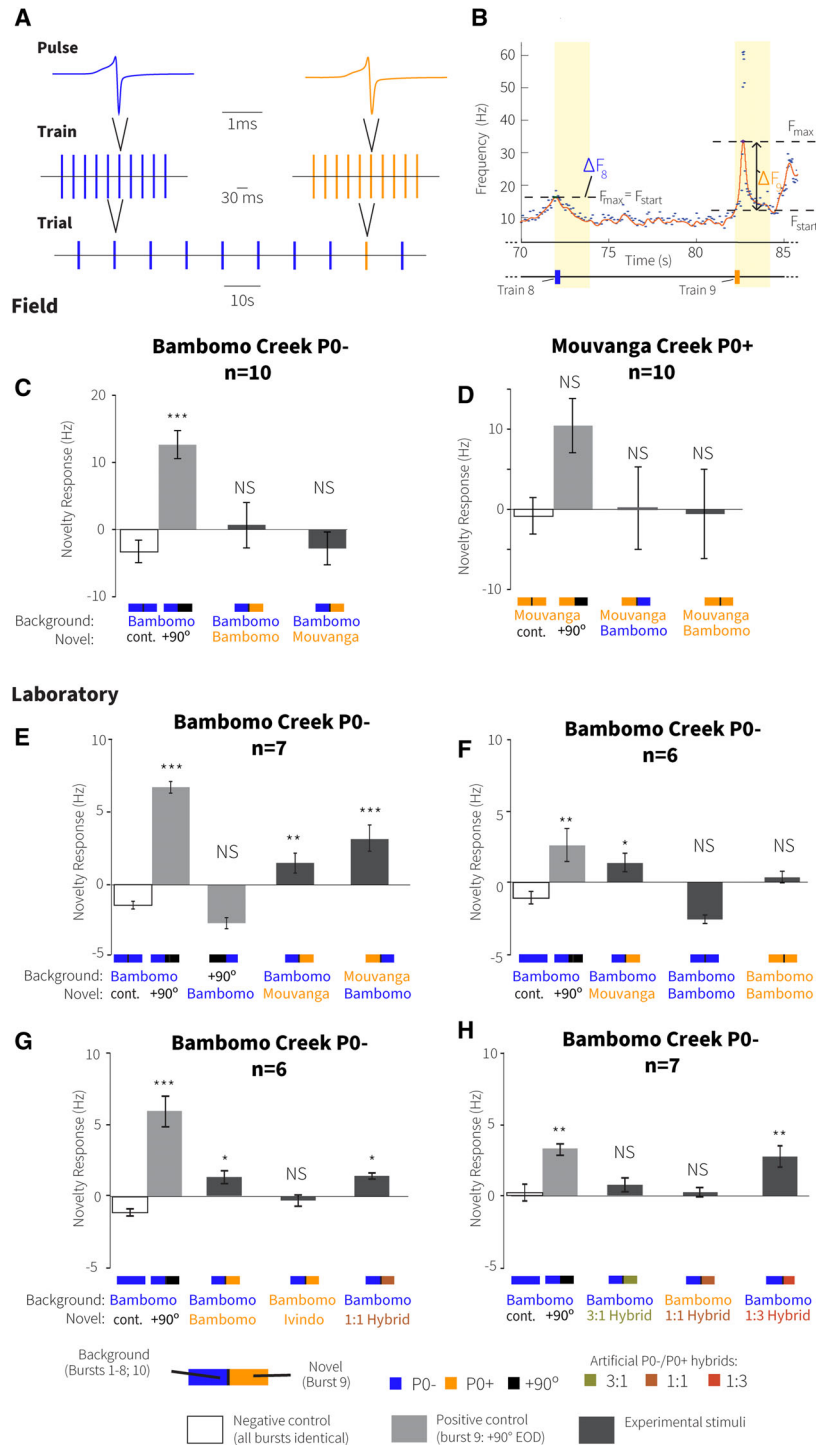


Figure 8. Results of dishabituation playback experiments performed on *P. kinsleyae*. **A.** Stimuli used: 10 trains of 10 EODs each are presented in each trial. 8 trains were comprised of one repeated EOD waveform type (background) followed by one train of 10 EODs (novel) followed by one train of 10 background EODs. **B.** Example of subject response. Subject's EOD discharges were continuously monitored (black dots), and discharge frequency was fitted with a spike density function (red line). The maximum and starting discharge rate of the fish was calculated over the 2sec interval from the onset of each train presentation. The difference between these values defined the change in frequency (ΔF) for that train. Novelty response was defined as the difference in ΔF between the 8th and 9th (novel) train presentation. See methods for details. **C-D.** Experiments performed on field captured specimens. **E-H.** Experiments performed in the laboratory. Localities and sample sizes are described above each plot. Bars show mean novelty responses averaged for all specimens $\pm$ standard error of the mean. Subtext indicates the localities where background and novel waveforms were recorded, if applicable. Stars reflect significance as compared to negative control (Dunnett's Test w/Control: NS: not significant, * $p=0.05$; ** $p<0.01$; *** $p<0.001$).

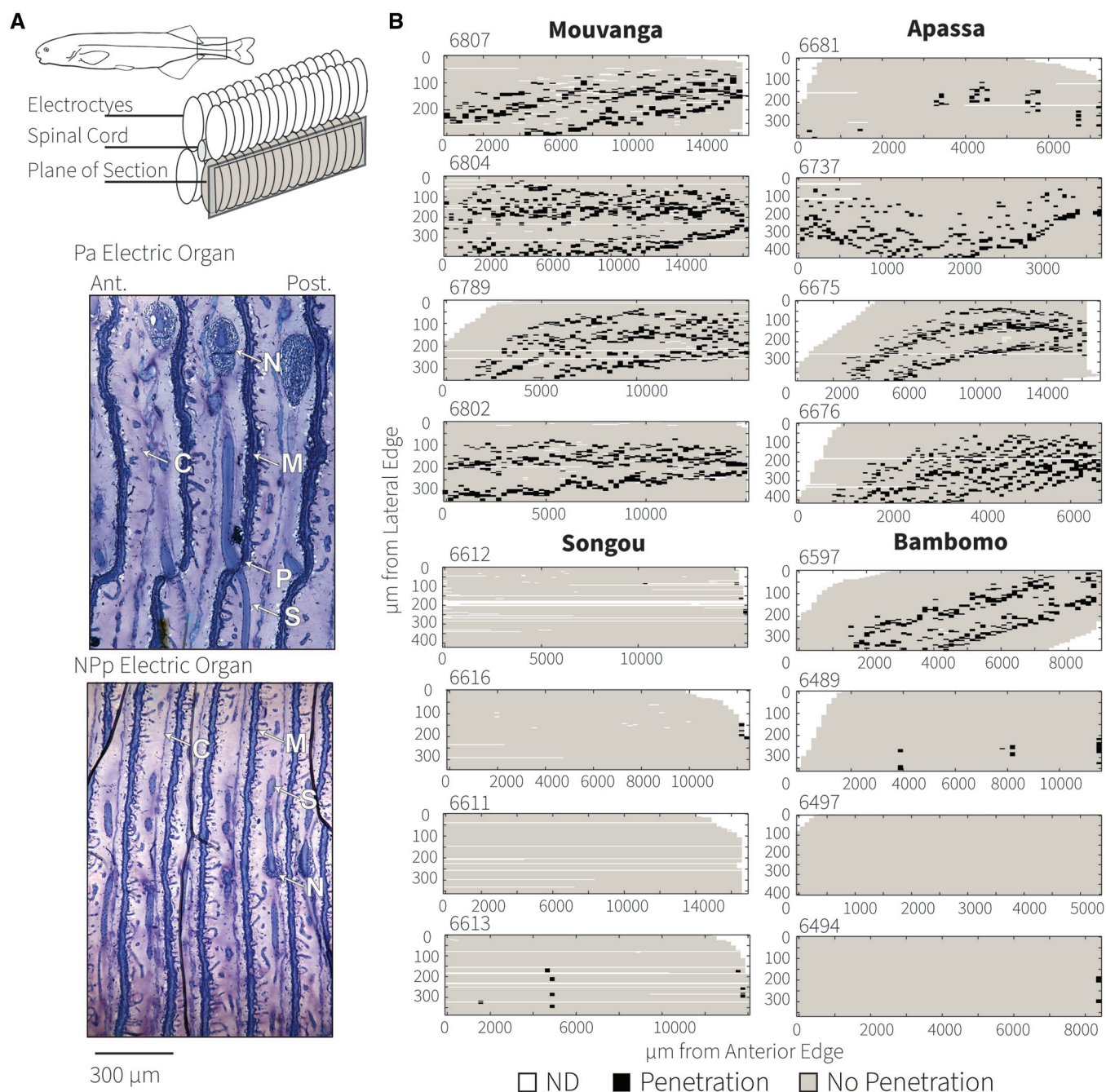


Figure 9. Summary of histological survey of electric organs, which are restricted to the caudal peduncle in mormyrids. **A.** Individual *P. kingsleyae* were sectioned sagittally from lateral to medial, following one of the four column of electrocytes that comprise the electric organ. Example histology showing the two basic types of anatomical configuration in the electric organ: *Pa*-type (stalk-penetrated with anterior innervation) and *NPP*-type (non-penetrated with posterior innervation). Stalks (S) can be seen clearly passing through the electrocyte penetrations (P) in *Pa* but not in *NPP* type electrocytes. Microstalklets (M) can be observed on the posterior face in both cases, and connective tissue boundaries (C) bounding each electrocyte are indicated. Innervation (N) can be observed on the anterior or posterior side of the electrocyte accordingly. *Pa*-type electrocytes result in EOD signals that have a small head negative phase P₀, which is absent in individuals with electrocytes that have *NPP* electrocyte anatomy (see Gallant et al. (2011)). **B.** Examples of histological analysis of four electric organs from each population. Each pixel represents an individual electrocyte in an individual section that was scored for presence of penetrations (black). In Apassa creek, most individuals have *Pa*-type observed in each electrocyte from anterior to posterior, whereas one individual had *NPP*-type and *Pa*-type electrocytes (patches of black in mostly grey background). In Bambomo, most individuals were *NPP* with one individual exhibiting all *Pa* type electrocytes. Songou was entirely comprised of individuals with *NPP*-type electrocytes, and Mouvanga was comprised of individuals with entirely *Pa*-type electrocytes. Full analysis is summarized in Table 4.

likely related to the balance between mutational processes within populations and gene flow between them (Hutchison and Templeton 1999), whereby gene flow and drift influence regional population structure differently depending on scale. As microsatellites have very high mutation rates and unique mutational processes (Selkoe and Toonen 2006), it is likely that the signal of gene flow between Southern and Northern populations is overwhelmed by the influence of genetic drift and mutation, thus reducing F_{st} estimates. Future studies, with more numerous markers (i.e., single nucleotide polymorphisms), will be necessary to fully understand *P. kingsleyae*'s genetic structure and colonization history over the Gabonese landscape. Third, we note that in Bambomo and Mouvanga Creeks, allele frequencies were remarkably stable over a 10-year period, suggesting large effective population sizes, at least in these locations (Waples 1989).

At a small geographic scale (i.e., Southern populations) where migration is much more likely between populations, the combination of strong genetic-geographic correlation with the total lack of correlation of EOD divergence with any variable (genetic or geographic) strongly implies that differences in selective pressures between populations are a likely important determinant of phenotypic divergence. At larger geographic scales, where migration is highly unlikely, the relationships between genetic, geographic, and phenotypic distances must be interpreted with more care for two reasons. First, microsatellite marker loci may underestimate genetic distances between geographically distant populations (as mentioned above). Second, the gap distribution in our sampling efforts between Southern and Northern populations may introduce potential confounding regional-level effects on EODs. Given these limitations, at this scale, our data identified that only PCA-derived signal and geographic distances were significantly correlated. This relationship disappeared when using MDS-derived signal distances and correcting for regional-level effects (Table 3). These results may be indicative of a potential clinal EOD variation pattern along geographic degrees of isolation such as reported in Gallant et al. (2011). However, the disparity between signal measures calls for caution and suggests that *P. kingsleyae*'s pattern of geographical EOD variation most likely does not simply result from classic isolation-by-distance processes (i.e., limited migration and gradual genetic drift). It also supports the likelihood of regional-level effects actively acting on phenotypic divergence, possibly in the form of different regional selective pressures or different regional colonization histories (e.g., founder effects). At these geographical scales, our results can speak to a relatively low likelihood of drift acting alone on signals. Nevertheless, studies should undoubtedly focus on a larger number of markers and a more continuous sampling scheme to corroborate these findings.

POTENTIAL SELECTIVE PRESSURES ON EOD DIVERSIFICATION

Ecological selection

Previous studies have considered ecological selection as an unlikely driver of EOD waveform divergence in the *Paramormyrops* radiation (Arnegard et al. 2010b; Gallant et al. 2011). However, studies on a second radiation of mormyrids in the *Campylomormyrus* genus have implicated ecological selection in the evolution of divergent snout morphologies and EOD types (Feulner et al. 2009b). Low-capacitance objects such as small invertebrate larvae, which seem to be a major part of mormyrid diets (Blake 1977; Hyslop 1986; Nwani et al. 2006), attenuate higher frequencies more readily (Meyer 1982; von der Emde and Ringer 1992; Crampton 1998). Recent work on mormyrid electrolocation has revealed that animals, plants, and invertebrates all create unique amplitude and waveform modulations referred to “electric colors” that allow these animals to reliably identify and differentiate these items (Gottwald et al. 2018), indicating that different EOD waveforms could therefore differ in their range of “electric color” detections.

In our study, of the three frequency-content variables measured, we found significant differences only in the high-frequency content of EOD signals (expressed by the variable *ffthi*, frequency above the peak power frequency at -3 dB) between triphasic and biphasic individuals (mean *ffthi* for triphasic individuals = 2200.3 Hz; mean *ffthi* for biphasic individuals = 2854.6 Hz; t -test $t = 8.32$; $P < 0.001$). It is therefore conceivable that biphasic versus triphasic EODs in *P. kingsleyae* may confer differential electrolocation capacities, particularly for prey detection, a topic which will be investigated in future studies.

Assortative mating

A communication context where EODs constitute the primary communication channel for mormyrids is sexual signaling, where males and females actively interact with one another (Bratton and Kramer 1989; Werneyer and Kramer 2005; Wong and Hopkins 2007). More specifically, many experimental studies have shown that both males and females exhibit mating preferences for specific EOD waveform features (Hopkins and Bass 1981; Arnegard et al. 2006; Machnik and Kramer 2008; Markowski et al. 2008; Feulner et al. 2009a; Machnik et al. 2010; Nagel et al. 2018a,b). Among *Paramormyrops* species, field and laboratory playback experiments have also revealed strong preferences for species-specific EOD waveforms during courtship (Hopkins and Bass 1981), indicating that EODs are most likely involved in maintaining prezygotic isolation between closely related species.

Our data are consistent with some degree of assortative mating between biphasic and triphasic *P. kingsleyae* EOD

Table 4. Summary of electric organ histological survey, performed on 21 specimens of *Paramormyrops kingsleyae*.

Population	Specimen No.	μm Surveyed	No. Sections	No. Electrocytes	No. <i>Pa</i>	No. <i>NPP</i>	Morphology
Apassa	6681	420	70	60	15	45	Mixed
Apassa	6675	648	108	64	50	14	Mixed
Apassa	6676	366	61	69	58	11	<i>Pa</i>
Apassa	6637	258	43	70	65	5	<i>Pa</i>
Bambomo	6489	240	40	60	10	50	<i>NPP</i>
Bambomo	6494	384	64	61	1	60	<i>NPP</i>
Bambomo	6497	276	46	67	0	67	<i>NPP</i>
Bambomo	6500	270	45	59	1	58	<i>NPP</i>
Bambomo	6547	378	63	52	1	51	<i>NPP</i>
Bambomo	6549	186	31	60	5	55	<i>NPP</i>
Bambomo	6597	438	73	58	43	15	Mixed
Mouvanga	6789	516	86	58	53	5	<i>Pa</i>
Mouvanga	6811	342	57	62	55	7	<i>Pa</i>
Mouvanga	6807	582	97	50	49	1	<i>Pa</i>
Mouvanga	6810	390	65	66	61	5	<i>Pa</i>
Mouvanga	6804	570	95	65	65	0	<i>Pa</i>
Mouvanga	6802	570	95	57	57	0	<i>Pa</i>
Songou	6611	252	42	59	0	59	<i>NPP</i>
Songou	6612	564	94	70	4	66	<i>NPP</i>
Songou	6613	234	39	64	5	59	<i>NPP</i>
Songou	6616	480	80	57	2	55	<i>NPP</i>

For each individual, the collection locality and specimen number are provided. For each specimen, one column of the entire electric organ was surveyed for the specified depth (μm) from lateral to medial (see Fig. 9). Total number of electrocytes and the number of those exhibiting *Pa* type morphology and *NPP* type morphology are provided. Finally, the assessment of the overall EO morphology is specified as either *Pa* (>75% of electrocytes *Pa*), *NPP* (>75% electrocytes *NPP*), or mixed (<75% of electrocytes of one type). Note that some individuals determined to have *Pa* anatomy did not have penetrations in the rostral or caudal portion of the organ (see Fig. 9). In these cases, electrocytes could not be fully surveyed because of the orientation of tissue during sectioning.

types. However, it is likely incomplete given numerous lines of evidence for hybridization. First, we found weak yet significant genetic differentiation between Apassa and Bambomo Creeks ($F_{st} < 0.07$), despite the fact that these populations are nearly fixed for alternate electric organ anatomies and signal types. Another line of evidence is that both populations exhibit a so-called “rare alleles” phenomenon (Fig. 5): both have additional alleles that are absent from all other populations genotyped, which has been demonstrated as a signature of hybrid zones in other taxa (Golding 1983; Barton 1985; Hoffman and Brown 1995). A third line of evidence is the existence of mixed electric organ morphology individuals in both Apassa and Bambomo (Fig. 9 and Table 4 in addition to those already discovered in Gallant et al. [2011]). We note that the only other documented case of mormyrids with mixed-morphology electrocytes was described in artificially created crosses between the biphasic mormyrid *Campylomormyrus tshokwe* and triphasic *Campylomormyrus tamandua* (Kirschbaum et al. 2016).

Our behavioral experiments provide evidence that *P. kingsleyae* from the putative hybrid zone are capable of discriminating

between P0-present and P0-absent EODs, a finding that is consistent with several studies that have characterized the neural encoding of EOD waveforms (Amagai et al. 1998; Friedman and Hopkins 1998; Xu-Friedman and Hopkins 1999; Carlson 2009; Carlson and Arnegard 2011; Baker et al. 2013; Lyons-Warren et al. 2013). These results indicate that in zones of signal sympatry, the observed EOD variation has the potential to be meaningful to *P. kingsleyae* receivers and could be a basis for assortative mating in this species. It is possible that hybridization is facilitated by the considerable variation in discrimination ability between individuals that we found in our field and laboratory experiments, despite the overall evidence of discrimination (Fig. 8). Variation in discrimination and perceptual biases among choosers in the context of mate choice has been demonstrated extensively (Rodríguez and Andrew Snedden 2004; Ryan and Cummings 2013) and can have important implications for hybridization and evolutionary diversification. This variation may accurately reflect that not all *P. kingsleyae* are equally good at discriminating EOD waveforms, which may be due to differences in reproductive state, age, condition, and sensory acuity of the individuals (Rosenthal 2017). In the same way, a lack of novelty

response in our experiments does not necessarily equate a lack of discrimination, as some fish may be able to distinguish signals and still not show a behavioral response.

We can conceive of two alternative scenarios that may explain the divergence of EOD signals in *P. kingsleyae*. First, *P. kingsleyae* populations could evolve divergence in EODs and concordant divergence in mate preferences due to sexual selection, which has been found to facilitate the rapid evolution of mating signals in many anurans (e.g., Pröhl et al. 2006; Boul et al. 2007; Pröhl et al. 2007; Lemmon 2009) and even to drive speciation in some of these cases (Masta and Maddison 2002; Boul et al. 2007). Second, variation in *P. kingsleyae* signals may be due to reinforcing selection against maladaptive hybridization between populations that initially diverged due to natural selection on ecological traits (Dobzhansky 1937).

Under both scenarios, we would expect *P. kingsleyae* to exhibit preferences for local versus foreign EODs in *P. kingsleyae*, which is an important future study. Second, the likelihood of these alternative scenarios is largely contingent on assessing the relative fitness and frequency of hybrid individuals. The small proportion of intermediate morphology individuals did not show obvious signs of morphological or behavioral abnormalities. However, characterizing the relative fitness of these individuals compared to phenotypically normal individuals is another important area of future research. Plausibly, if differences in EOD are adaptations to capturing different prey items (see preceding section), hybrid EOD signals could perform relatively poorly in prey detection. This would suggest that the presence or absence of P0 might act similarly to a “magic trait” (Feulner et al. 2009b; Gavrillets 2004), that is, a trait resulting from divergent ecological selection that also contributes to assortative mating.

Concluding Remarks

Our study demonstrates that drift is insufficient to explain EOD diversity within *P. kingsleyae* and supports an important role of selection in the evolution of EODs, even on very small geographic scales within a single species. Although drift has been found to play an important role in the divergence of communication signals in several taxa including South American electric fish (Picq et al. 2016), Neotropical singing mice (Campbell et al. 2010), Amazonian and Microhylid frogs (Amézquita et al. 2009; Lee et al. 2016), and greenish warblers (Irwin et al. 2008), it is important to note that most studies testing the contribution of drift in communication signals rarely report evidence for neutral signal evolution (Soha et al. 2003; Nicholls et al. 2006; Pröhl et al. 2006; Ruegg et al. 2006; Pröhl et al. 2007; Rudh et al. 2007; Dingler et al. 2008; Huttunen et al. 2008; Tobias et al. 2010; Cadena et al. 2011; Lin et al. 2015; Sathyan et al. 2017).

Last, signal evolution is rarely explained by a single force; drift has been implicated to work in conjunction with selection in driving signal divergence both theoretically (Uyeda et al. 2009) and empirically in many signaling modalities including echolocation calls (Jacobs and Mutumi 2018), bird songs (Irwin et al. 2008), mice songs (Campbell et al. 2010), and electric fish signals (Picq et al. 2016).

The intraspecific polymorphism in the signals of *P. kingsleyae* is recurrent between *Paramormyrops* species, a common pattern in species radiations, which gives the impression that certain traits “blink on and off” during evolution (West-Eberhard 2003). These parallelisms inform us on how key traits, in this case EODs, can initially diverge and diversify. Our results suggest that divergent selection operating over small geographic scales promotes EOD signal divergence for a widely variable character in mormyrid electric fish. This phenomenon is consistent with other species radiations characterized by intra- and interspecific parallelisms in key traits, where either sexual selection (e.g., in East African cichlids [Maan et al. 2004] and in coral reef hamlet fish [Puebla et al. 2012]) or natural selection (e.g., in Darwin’s finches [Werner and Sherry 1987]) have been identified as the main drivers of diversification. The framework provided by this study presents a clear set of testable hypotheses, such that future studies might explicitly identify the source of selection acting on EOD signals in *P. kingsleyae*. These may be the same forces that contributed to the rapid radiation of the *Paramormyrops* genus, and of other mormyrid genera.

AUTHOR CONTRIBUTIONS

SP performed signal and genetic variation analyses and co-designed the study. CC performed histological analyses. JS performed laboratory discrimination experiments. BAC performed field discrimination experiments and co-designed the study. JRG co-designed the study, collected specimens, performed microsatellite genotyping, and oversaw analysis of EODs and population genetics. All authors contributed to writing the manuscript.

ACKNOWLEDGMENTS

Permits to collect fishes in Gabon and export them for this study were granted by l’Institut de Recherche en Ecologie Tropicale, l’Institut de Recherches Agronomiques et Forestières, and the Centre National de la Recherche Scientifique et Technologique. We are grateful for the valuable assistance and logistical support we received from J. D. Mbega and students working in these institutions. All techniques used are in accordance with protocols approved by Cornell University’s Center for Research Animal Resources and Education (CARE). Additionally, we thank C. D. Hopkins (CDH), M. Arnegard, A. McCune, K. Shaw, J. Fetcho, D. Deitcher, and S. Mullen as well as two anonymous reviewers for comments on earlier versions of this manuscript. This work was supported by NIMH TG T32 MH015793, NIH TG 2T32GM007469, and National Science Foundation 1455405 to JRG, NIH RO1-DC6206, National Science Foundation 0818305 to CDH, and National Science Foundation 0818390 as well as National Science Foundation 1255396 to BAC.

DATA ARCHIVING

Source code for this analysis is available on GitHub (<https://doi.org/10.5281/zenodo.3691905>) and all original data are archived in Dryad (<https://doi.org/10.5061/dryad.2z34tmphj>).

LITERATURE CITED

- Albert, J., and R. Reis. 2011. Introduction to Neotropical freshwaters. Pp. 3–19 in J. S. Albert and R. Reis, eds. *Historical biogeography of Neotropical freshwater fishes*. Univ. of California Press, Los Angeles, CA.
- Allender, C. J., O. Seehausen, M. E. Knight, G. F. Turner, and N. Maclean. 2003. Divergent selection during speciation of Lake Malawi cichlid fishes inferred from parallel radiations in nuptial coloration. *Proc. Natl Acad. Sci.* 100:14074–14079.
- Amagai, S., M. Friedman, and C. Hopkins. 1998. Time coding in the midbrain of mormyrid electric fish. I. Physiology and anatomy of cells in the nucleus extero-lateralis pars anterior. *J. Comp. Physiol. A-Neuroethol. Sens. Neural Behav. Physiol.* 182:115–130.
- Amézquita, A., A. P. Lima, R. Jelhe, L. Castellanos, Ó. Ramos, A. J. Crawford, H. Gasser, and W. Hödl. 2009. Calls, colours, shape, and genes: a multi-trait approach to the study of geographic variation in the Amazonian frog *Allobates femoralis*. *Biol. J. Linn. Soc.* 98:826–838.
- Arnegard, M. E., and C. D. Hopkins. 2003. Multivariate analysis of electric signal variation among seven blunt-snouted Brienomyrus species (Teleostei: Mormyridae) from Gabon. *Environ. Biol. Fishes* 67:321–339.
- Arnegard, M. E., S. M. Bogdanowicz, and C. D. Hopkins. 2005. Multiple cases of striking genetic similarity between alternate electric fish signal morphs in sympatry. *Evolution* 59:324–343.
- Arnegard, M. E., B. S. Jackson, and C. D. Hopkins. 2006. Time-domain signal divergence and discrimination without receptor modification in sympatric morphs of electric fishes. *J. Exp. Biol.* 209:2182–2198.
- Arnegard, M. E., D. Zwickl, and Y. Lu. 2010a. Old gene duplication facilitates origin and diversification of an innovative communication system—twice. *Proc. Natl. Acad. Sci. USA* 107:22172–22177.
- Arnegard, M. E., P. B. McIntyre, L. J. Harmon, M. L. Zelditch, W. G. R. Crampton, J. K. Davis, J. P. Sullivan, S. Lavoué, and C. D. Hopkins. 2010b. Sexual signal evolution outpaces ecological divergence during electric fish species radiation. *Am. Nat.* 176:335–356.
- Baker, C., T. Kohashi, A. Lyons-Warren, X. Ma, and B. Carlson. 2013. Multiplexed temporal coding of electric communication signals in mormyrid fishes. *J. Exp. Biol.* 216:2365–2379.
- Barton, N. 1985. Analysis of hybrid zones. *Annu. Rev. Ecol. Syst.* 16:113–148.
- Bennett, M. V. L., and H. Grundfest. 1961. Studies on the morphology and electrophysiology of electric organs III. Electrophysiology of electric organs in mormyrids. Pp. 113–135 in C. Chagas and A. Paes de Carvalho, eds. *Bioelectrogenesis*. Elsevier, Amsterdam, the Netherlands.
- Blake, B. F. 1977. Food and feeding of the mormyrid fishes of Lake Kainji, Nigeria, with special reference to seasonal variation and interspecific differences. *J. Fish Biol.* 11:315–328.
- Boul, K. E., W. Chris Funk, C. R. Darst, D. C. Cannatella, and M. J. Ryan. 2006. Sexual selection drives speciation in an Amazonian frog. *Proc. R. Soc. B: Biol. Sci.* 274:399–406.
- . 2007. Sexual selection drives speciation in an Amazonian frog. *Proc. R. Soc. B: Biol. Sci.* 274:399–406.
- Bratton, B. O., and B. Kramer. 1989. Patterns of the electric organ discharge during courtship and spawning in the mormyrid fish, *Pollimyrus isidori*. *Behav. Ecol. Sociobiol.* 24:349–368.
- Brower, A. V. Z. 1994. Rapid morphological radiation and convergence among races of the butterfly *Heliconius erato* inferred from patterns of mitochondrial DNA evolution. *Proc. Natl. Acad. Sci.* 91:6491–6495.
- Cadena, C. D., Z. A. Cheverson, and W. C. Funk. 2011. Testing the molecular and evolutionary causes of a ‘leapfrog’ pattern of geographical variation in coloration. *J. Evol. Biol.* 24:402–414.
- Campbell, P., B. Pasch, J. L. Pino, O. L. Crino, M. Phillips, and S. M. Phelps. 2010. Geographic variation in the songs of Neotropical singing mice: testing the relative importance of drift and local adaptation. *Evolution* 64:1955–1972.
- Carlson, B. A. 2009. Temporal-pattern recognition by single neurons in a sensory pathway devoted to social communication behavior. *J. Neurosci.* 29:9417–9428.
- Carlson, B. A., and C. D. Hopkins. 2004. Stereotyped temporal patterns in electrical communication. *Anim. Behav.* 68:867–878.
- Carlson, B. A., and J. R. Gallant. 2013. From sequence to spike to spark: evo-devo-neuroethology of electric communication in mormyrid fishes. *J. Neurogenet.* 27:106–129.
- Carlson, B. A., and M. E. Arnegard. 2011. Neural innovations and the diversification of African weakly electric fishes. *Commun. Integr. Biol.* 4:720–725.
- Carlson, B.A., Hasan, S.M., Hollmann, M., Miller, D.B., Harmon, L.J., and Arnegard, M.E. 2011. Brain evolution triggers increased diversification of electric fishes. *Science* 332(): 583–586.
- Coyne, J.A., and Orr, H.A. 2004. *Speciation*. Sunderland, MA: Sinauer Associates.
- Clarke, K. 1993. Non-parametric multivariate analyses of changes in community structure. *Australian Journal of Ecology* 18 117–143.
- Crampton, W. G. R. 1998. Electric signal design and habitat preferences in a species rich assemblage of gymnotiform fishes from the Upper Amazon Basin. *An.-Acad. Bras. Cienc.* 70:805–848.
- Diamond, J. 1986. Biology of birds of paradise and bowerbirds. *Annu. Rev. Ecol. Syst.* 17:17–37.
- Dingle, C., W. Halfwerk, and H. Slabbekoorn. 2008. Habitat-dependent song divergence at subspecies level in the grey-breasted wood-wren. *J. Evol. Biol.* 21:1079–1089.
- Dobzhansky, T. 1937. *Genetics and the origin of species*. New York, NY: Columbia University Press.
- Dunnnett, C. W. 1955. A multiple comparison procedure for comparing several treatments with a control. *J. Am. Statist. Assoc.* 50:1096–1121.
- Excoffier, L., and H. E. L. Lischer. 2010. Arlequin suite ver 3.5: a new series of programs to perform population genetics analyses under Linux and Windows. *Mol. Ecol. Resour.* 10:564–567.
- Feulner, P. G., M. Plath, J. Engelmann, F. Kirschbaum, and R. Tiedemann. 2009a. Electrifying love: electric fish use species-specific discharge for mate recognition. *Biol. Lett.* 5:225–228.
- . 2009b. Magic trait electric organ discharge (EOD). *Commun. Integr. Biol.* 2:329–331.
- Friedman, M. A., and C. D. Hopkins. 1998. Neural substrates for species recognition in the time-coding electrosensory pathway of mormyrid electric fish. *J. Neurosci.* 18:1171–1185.
- Gallant, J. R., M. E. Arnegard, J. P. Sullivan, B. A. Carlson, and C. D. Hopkins. 2011. Signal variation and its morphological correlates in *Paramormyrops kingsleyae* provide insight into evolution of electrogenic signal diversity in mormyrid fish. *J. Comp. Physiol. A* 197:799–817.
- Gavrilets, S. 2004. *Fitness landscapes and the origin of species*. Princeton Univ. Press, Princeton, NJ.
- Glaubrecht, M. 2010. *Evolution in action: case studies in adaptive radiation, speciation and the origin of biodiversity*. Springer, New York.

- Golding, G. 1983. Increased number of alleles found in hybrid populations due to intragenic recombination. *Evolution* 37:1–17.
- Gottwald, M., N. Singh, A. N. Haubrich, S. Regett, and G. von der Emde. 2018. Electric-color sensing in weakly electric fish suggests color perception as a sensory concept beyond vision. *Curr. Biol.* 28:3648–3653.
- Heiligenberg, W. 1977. Principles of electrolocation and jamming avoidance in electric fish: a neuroethological approach. Springer, New York.
- Heiligenberg, W., and R. A. Altes. 1978. Phase sensitivity in electroreception. *Science* 199:1001–1004.
- Hoffman, S. M. G., and W. M. Brown. 1995. The molecular mechanism underlying the “rare allele phenomenon” in a subspecific hybrid zone of the California field mouse, *Peromyscus californicus*. *J. Mol. Evol.* 41:1165–1169.
- Hopkins, C. D., and A. Bass. 1981. Temporal coding of species recognition signals in an electric fish. *Science* 212:85–87.
- Hutchison, D., and A. Templeton. 1999. Correlation of pairwise genetic and geographic distance measures: inferring the relative influences of gene flow and drift on the distribution of genetic variability. *Evolution* 53:1898–1914.
- Huttunen, S., J. Aspi, C. Schlotterer, J. Routtu, and A. Hoikkala. 2008. Variation in male courtship song traits in *Drosophila virilis*: the effects of selection and drift on song divergence at the intraspecific level. *Behav. Genet.* 38:82–92.
- Hyslop, E. J. 1986. The food habits of four small-sized species of Mormyridae from the floodplain pools of the Sokoto-Rima river basin, Nigeria. *J. Fish Biol.* 28:147–151.
- Irwin, D. E., M. P. Thimman, and J. H. Irwin. 2008. Call divergence is correlated with geographic and genetic distance in greenish warblers (*Phylloscopus trochiloides*): a strong role for stochasticity in signal evolution? *J. Evol. Biol.* 21:435–448.
- Jacobs, D. S., and G. L. Mutumi. 2018. The relative roles of selection and drift in phenotypic variation: some like it hot, some like it wet. Pp. 215–237 in P. Pontarotti, ed. *Origin and evolution of biodiversity*. Springer, Cham, Switzerland.
- Kirkpatrick, M., and M. J. Ryan. 1991. The evolution of mating preferences and the paradox of the lek. *Nature* 350:33–38.
- Kirschbaum, F., L. Nguyen, S. Baumgartner, H. W. L. Chi, R. Wolfart, K. Elarabani, H. Eppenstein, Y. Kornienko, L. Guido-Bohm, V. Mamonekene, et al. 2016. Intra-genus (*Campylomormyrus*) and inter-genus hybrids in mormyrid fish: physiological and histological investigations of the electric organ ontogeny. *J. Physiol.* 110:281–301.
- Kramer, B., and G. W. M. Westby. 1985. No sex difference in the waveform of the pulse type electric fish, *Gnathonemus petersii* (Mormyridae). *Experientia* 41:1530–1531.
- Kruskal, J., and M. Wish. 1978. *Multidimensional scaling*. Sage, Beverly Hills, CA.
- Lavoué, S., J. P. Sullivan, M. E. Arnegard, and C. D. Hopkins. 2008. Differentiation of morphology, genetics and electric signals in a region of sympatry between sister species of African electric fish (Mormyridae). *J. Evol. Biol.* 21:1030–1045.
- Lee, K. H., P. J. Shaner, Y. P. Lin, and S. M. Lin. 2016. Geographic variation in advertisement calls of a *Microhylid* frog—testing the role of drift and ecology. *Ecol. Evol.* 6:3289–3298.
- Lemmon, E. M. 2009. Diversification of conspecific signals in sympatry: geographic overlap drives multidimensional reproductive character displacement in frogs. *Evolution* 63:1155–1170.
- Liem, K. F., and L. S. Kaufmann. 1984. Intraspecific macroevolution: functional biology of the polymorphic cichlid species *Cichlasoma minckleyi*. Pp. 203–215 in A. A. Echelle and I. Kornfield, eds. *Evolution of fish species flocks*. Univ. of Maine Press, Orono, ME.
- Lin, A., T. Jiang, J. S. Kanwal, G. Lu, J. Luo, X. Wei, B. Luo, and J. Feng. 2015. Geographical variation in echolocation vocalizations of the Himalayan leaf-nosed bat: contribution of morphological variation and cultural drift. *Oikos* 124:364–371.
- Lissmann, H. W. 1958. On the function and evolution of electric organs in fish. *J. Exp. Biol.* 35:156–191.
- Lyons-Warren, A., T. Kohashi, S. Mennerick, and B. Carlson. 2013. Detection of submillisecond spike timing differences based on delay-line anti-coincidence detection. *J. Neurophysiol.* 110:2295–2311.
- Maan, M. E., O. Seehausen, L. Soderberg, L. Johnson, E. A. Ripmeester, H. D. Mrosso, M. I. Taylor, T. J. van Dooren, and J. J. van Alphen. 2004. Intraspecific sexual selection on a speciation trait, male coloration, in the Lake Victoria cichlid *Pundamilia nyererei*. *Proc. Biol. Sci.* 271:2445–2452.
- Machnik, P., and B. Kramer. 2008. Female choice by electric pulse duration: attractiveness of the males’ communication signal assessed by female bulldog fish, *Marcusenius pongolensis* (Mormyridae, Teleostei). *J. Exp. Biol.* 211:1969–1977.
- Machnik, P., B. Markowski, and B. Kramer. 2010. Intra- versus inter-sexual selection in the dimorphic electric organ discharges of the snout-fish *Marcusenius altisambesi* (Mormyridae, Teleostei). *Behaviour* 147:677–704.
- Mantel, N. 1967. The detection of disease clustering and a generalized regression approach. *Cancer Research* 27:209–220.
- Markowski, B., B. Baier, and B. Kramer. 2008. Differentiation in electrical pulse waveforms in a pair of sibling dwarf stonebushers, *Polimyrus castelnaui* and *P. marianne*: possible mechanisms and functions (Mormyridae, Teleostei). *Behaviour* 145:115–135.
- Masta, S. E., and W. P. Maddison. 2002. Sexual selection driving diversification in jumping spiders. *Proc. Natl. Acad. Sci.* 99:4442–4447.
- Mayr, E. 1963. *Animal species and evolution*. Cambridge, MA: Belknap Press.
- Meirmans, P. G. 2012. The trouble with isolation by distance. *Mol. Ecol.* 21:2839–2846.
- Mendelson, T. C., and K. L. Shaw. 2005. Rapid speciation in an arthropod. *Nature* 433:375–376.
- Meyer, A. 1987. Phenotypic plasticity and heterochrony in *Cichlasoma managuense* (Pisces, Cichlidae) and their implications for speciation in cichlid fishes. *Evolution* 41:1357–1369.
- Meyer, H. 1982. Behavioral responses of weakly electric fish to complex impedances. *J. Comp. Physiol.* 145:459–470.
- Mullen, S. P., T. C. Mendelson, C. Schal, and K. L. Shaw. 2007. Rapid evolution of cuticular hydrocarbons in a species radiation of acoustically diverse Hawaiian crickets (Gryllidae: Trigonidiinae: Laupala). *Evolution* 61:223–231.
- Nagel, R., F. Kirschbaum, J. Engelmann, V. Hofmann, F. Pawelzik, and R. Tiedemann. 2018a. Male-mediated species recognition among African weakly electric fishes. *R. Soc. Open Sci.* 5:170443.
- Nagel, R., F. Kirschbaum, V. Hofmann, J. Engelmann, and R. Tiedemann. 2018b. Electric pulse characteristics can enable species recognition in African weakly electric fish species. *Sci. Rep.* 8:10799.
- Nicholls, J. A., C. Austin, C. Moritz, and A. W. Goldizen. 2006. Genetic population structure and call variation in a passerine bird, the satin bowerbird, *Ptilonorhynchus violaceus*. *Evolution* 60:1279–1290.
- Nwani, C. D., J. E. Eyo, and E. F. Udeh. 2006. Food and feeding habits of *Campylomormyrus tamandua* in Anambra river, Nigeria. *Anim. Res. Int.* 3:410–414.

- Oksanen, J., F. G. Blanchet, F. M. R. Kindt, P. Legendre, D. McGlinn, P. R. Minchin, R. B. O'Hara, G. L. Simpson, P. Solymos, et al. 2018. *vegan*: community ecology package. R package version 2.4-6.
- Picq, S., F. Alda, E. Bermingham, and R. Krahe. 2016. Drift-driven evolution of electric signals in a Neotropical knife-fish. *Evolution* 70:2134–2144.
- Picq, S., M. Scotti, and O. Puebla. 2019. Behavioural syndromes as a link between ecology and mate choice: a field study in a reef fish population. *Anim. Behav.* 150:219–237.
- Pröhl, H., S. Hagemann, J. Karsch, and G. Höbel. 2007. Geographic variation in male sexual signals in strawberry poison frogs (*Dendrobates pumilio*). *Ethology* 113:825–837.
- Pröhl, H., R. A. Koshy, U. Mueller, M. J. Rand, and M. J. Ryan. 2006. Geographic variation of genetic and behavioral traits in Northern and Southern túngara frogs. *Evolution* 60:1669–1679.
- Puebla, O., E. Bermingham, and F. Guichard. 2012. Pairing dynamics and the origin of species. *Proc. R. Soc. B: Biol. Sci.* 279:1085–1092.
- R Core Team. 2017. R: A language and environment for statistical computing. R Foundation for Statistical Computing, Vienna, Austria.
- Rodríguez, R. L., and W. Andrew Snedden. 2004. On the functional design of mate preferences and receiver biases. *Anim. Behav.* 68: 427–432.
- Rosenthal, G. G. 2017. *Mate choice: the evolution of sexual decision making from microbes to humans*. Princeton Univ. Press, Princeton, NJ.
- Rousset, F. 1997. Genetic differentiation and estimation of gene flow from F-statistics under isolation by distance. *Genetics* 145 1219–1228.
- Rousset, F. 2008. GENEPOP'007: a complete re-implementation of the GENEPOP software for windows and linux. *Mol. Ecol. Resour.* 8:103–106.
- Rudh, A., B. Rogell, and J. Hoglund. 2007. Non-gradual variation in colour morphs of the strawberry poison frog *Dendrobates pumilio*: genetic and geographical isolation suggest a role for selection in maintaining polymorphism. *Mol. Ecol.* 16:4284–4294.
- Ruegg, K., H. Slabbekoorn, S. Clegg, and T. B. Smith. 2006. Divergence in mating signals correlates with ecological variation in the migratory songbird, Swainson's thrush (*Catharus ustulatus*). *Mol. Ecol.* 15:3147–3156.
- Ryan, M. J., and M. E. Cummings. 2013. Perceptual biases and mate choice. *Annu. Rev. Ecol. Evol. Syst.* 44:437–459.
- Ryman, N., and S. Palm. 2006. POWSIM: a computer program for assessing statistical power when testing for genetic differentiation. *Mol. Ecol.* 6:600–602.
- Sathyan, R., A. Engelbrecht, and V. C. K. Couldridge. 2017. Morphological, acoustic and genetic divergence in the bladder grasshopper *Bullacris unicolor*. *Ethol. Ecol. Evol.* 29:552–573.
- Schluter, D., and J. D. McPhail. 1992. Ecological character displacement and speciation in sticklebacks. *Am. Nat.* 140:85–108.
- Selkoe, K. A., and R. J. Toonen. 2006. Microsatellites for ecologists: a practical guide to using and evaluating microsatellite markers. *Ecol. Lett.* 9:615–629.
- Smith, T. B. 1987. Bill size polymorphism and intraspecific niche utilization in an African finch. *Nature* 329:717–719.
- Smouse, P. E., J. C. Long, and R. R. Sokal. 1986. Multiple regression and correlation extensions of the Mantel test of matrix correspondence. *Syst. Zool.* 35:627–632.
- Soha, J. A., D. A. Nelson, and P. G. Parker. 2003. Genetic analysis of song dialect populations in Puget Sound white-crowned sparrows. *Behav. Ecol.* 15:636–646.
- Sullivan, J. P., S. Lavoué, M. E. Arnegard, and C. D. Hopkins. 2004. AFLPs resolve phylogeny and reveal mitochondrial introgression within a species flock of African electric fish (Mormyroidea: Teleostei). *Evolution* 58:824–841.
- Sullivan, J. P., S. Lavoue, and C. D. Hopkins. 2000. Molecular systematics of the African electric fishes (Mormyroidea: Teleostei) and a model for the evolution of their electric organs. *J. Exp. Biol.* 203:665–683.
- . 2002. Discovery and phylogenetic analysis of a riverine species flock of African electric fishes (Mormyridae: Teleostei). *Evolution* 56:597–616.
- Tobias, J. A., J. Aben, R. T. Brumfield, E. P. Derryberry, W. Halfwerk, H. Slabbekoorn, and N. Seddon. 2010. Song divergence by sensory drive in Amazonian birds. *Evolution* 64:2820–2829.
- Uyeda, J. C., S. J. Arnold, P. A. Hohenlohe, and L. S. Mead. 2009. Drift promotes speciation by sexual selection. *Evolution* 63:583–594.
- von der Emde, G., and T. Ringer. 1992. Electrolocation of capacitive objects in four species of pulse-type weakly electric fish I. Discrimination Performance. *Ethology* 91:326–338.
- Waples, R. S. 1989. A generalized approach for estimating effective population size from temporal changes in allele frequency. *Genetics* 121:379–391.
- Weir, B. S. 1990. *Genetic data analysis: methods for discrete population genetic data*. Sinauer, Sunderland, MA.
- Weir, B.S., and Cockerham, C.C. 1984. Estimating F-statistics for the analysis of population structure. *Evolution* 38(6): 1358–1370.
- Werner, T. K., and T. W. Sherry. 1987. Behavioral feeding specialization in *Pinaroloxias inornata*, the “Darwin's Finch” of Cocos Island, Costa Rica. *Proc. Natl Acad. Sci.* 84:5506–5510.
- Werner, M., and B. Kramer. 2005. Electric signalling and reproductive behaviour in a mormyrid fish, the bulldog *Marcusenius macrolepidotus* (South African form). *J. Ethol.* 23:113–125.
- West-Eberhard, M. J. 1986. Alternative adaptations, speciation, and phylogeny (A review). *Proc. Natl. Acad. Sci.* 83:1388–1392.
- West-Eberhard, M.J. 2003. *Developmental plasticity and evolution*, New York, NY: Oxford University Press.
- Wilkins, M.R., Seddon, N., and Safran, R.J. 2013. Evolutionary divergence in acoustic signals: causes and consequences. *Trends in Ecology & Evolution* 28(3): 156–166.
- Wong, R. Y., and C. D. Hopkins. 2007. Electrical and behavioral courtship displays in the mormyrid fish *Brienomyrus brachyistius*. *J. Exp. Biol.* 210:2244–2252.
- Wright, S. 1943. Isolation by distance. *Genetics* 28 139–156.
- Xu-Friedman, M., and C. Hopkins. 1999. Central mechanisms of temporal analysis in the knollenorgan pathway of mormyrid electric fish. *J. Exp. Biol.* 202:1311–1318.

Associate Editor: A. M. Rice
Handling Editor: M. R. Servedio

Supporting Information

Additional supporting information may be found online in the Supporting Information section at the end of the article.

Figure S1. Example Playback Waveforms: Hybrid EODs

Figure S2. Multidimensional scaling (MDS) plot of EOD waveform variation in 327 *P. kingsleyae* individuals from 9 populations in Gabon.

Figure S3. Comparison of inter-population signal distances estimated from PCA (Fig. 4) and from multidimensional scaling of cross-correlated waveforms (Fig. S2).

Figure S4. Mean (and range) of genetic distances for pairwise comparisons of Southern populations separated or on the same side of the Bongolo Falls.

Figure S5. Correlation plots between genetic, signal, and geographic distances for Southern populations only ($n = 8$ populations).

Table S2. Landmark and variable definitions.

Table S3. Top ten factor loading values for principal components 1 and 2.

Table S4. Results of standard and partial Mantel tests between genetic distances (F_{st}), signal distances (obtained through PCA (left) and MDS (right)), geographic distances (km), and presence/absence of Bongolo Falls (Southern populations) at two spatial scales.

Table S5. Results of standard and partial Mantel tests between genetic distances ($F_{st}/(1-F_{st})$), signal distances (obtained through PCA (left) and MDS (right)), geographic distances (km), and presence/absence of Bongolo Falls (Southern populations) at two spatial scales.

Table S6. Results of standard and partial Mantel tests between genetic distances (F_{st}), signal distances (obtained through PCA (left) and MDS (right)), geographic distances (km), and presence/absence of Bongolo Falls (Southern populations) at two spatial scales.

Figure A1. Multidimensional scaling (MDS) plot of Q_{10} corrected EOD waveform variation in 324 *P. kingsleyae* individuals from 11 populations in Gabon. Variation in waveforms was quantified through cross-correlation analysis of signals (see Carlson et al. (2011)).

Table A1. Results of standard and partial Mantel tests between genetic distances ($F_{st}/(1-F_{st})$), temperature corrected signal distances (obtained through MDS), geographic distances (km), and presence/absence of Bongolo Falls (Southern populations) at two spatial scales.

Figure A2. Power analysis simulations for obtaining significant outcomes in tests of genetic differentiation, using five microsatellite loci and the specific marker characteristics and sample sizes of this study.

Description of temperature correction on signal variation quantified through multidimensional scaling of signal cross correlations.

Description of power analysis of microsatellites for inferring genetic differentiation.